

1 **Title**

2 Hydrological regime and plant functional traits jointly mediate the influence of *Salix* spp. on  
3 soil organic carbon stocks in a High Arctic tundra

4 **Title shortened version**

5 Plant and environmental drivers of soil carbon stocks

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16 **Author contributions**

17 LJL, EL and VM conceived and designed the research. LJL, EL and MT collected the material.  
18 LJL and P-OC processed soil samples prior to analyses. LJL, EL, FG and VM supervised soil  
19 analyses. LJL, JFF, LD, VM and MG analyzed data. LJL and VM led the writing of the  
20 manuscript, with contributions of all co-authors.

## 21 **Abstract**

22 Evidence points out that increasing plant productivity associated with greater erect shrub  
23 abundance alters soil organic carbon (SOC) stocks in the Arctic. However, the underlying plant  
24 economic traits remain poorly examined, which limits our understanding of plant-environment  
25 interactions driving tundra carbon cycling. We explored how erect shrub abundance leads to  
26 SOC variation in a High Arctic tundra (Bylot Island, Nunavut, Canada), where the only erect  
27 shrub, *Salix richardsonii*, has settled along currently active and abandoned channel zones of  
28 alluvial fans. The effects of vegetation and local environmental changes on SOC were evaluated  
29 through a paired sampling of soil materials and plant aboveground functional traits associated  
30 with plant carbon supply and nutrient demand processes. The occurrence of *S. richardsonii*,  
31 characterized by a 10-fold increase in aboveground biomass, induced a 28% increase in SOC  
32 compared to adjacent plots dominated by prostrate shrubs and graminoids. Yet, this vegetation  
33 effect was solely observed along active channels, where higher SOC was associated with  
34 greater leaf and stem biomass. A path analysis showed that shrub leaf area index and total leaf  
35 nutrient content best represented plant carbon supply and nutrient demand dynamics,  
36 respectively, and jointly regulated SOC variation. This study underscores that vegetation  
37 structural changes associated with increasing erect shrub abundance in the Arctic can promote  
38 soil organic carbon storage, but that this pattern may be mediated by strong plant-environment  
39 interactions. Accounting for changes in functional traits driving plant carbon supply and  
40 nitrogen demand proves important for a better mechanistic understanding of how shrubification  
41 impacts tundra carbon cycling.

## 42 **Key words**

43 Alluvial fan; Carbon supply; Leaf area index; Nutrient demand; Plant-enhanced mineralization;  
44 Shrubification.

## 45 **Manuscript highlights**

46 - Increasing erect shrub abundance along High Arctic alluvial fan channels can lead to  
47 increasing soil organic carbon stocks

48 - Fine-scale interactions between hydrological regime and vegetation conditions regulate  
49 changes in soil organic carbon stocks

50 - Plant traits associated with carbon supply and nutrient demand may provide new insights on  
51 carbon cycling variation with vegetation shifts

## 52 **Introduction**

53 The greening of tundra ecosystems, mainly driven by the significant increase in erect shrub  
54 height (Bjorkman and others 2018), biomass and canopy cover (Myers-Smith and others 2011;  
55 Elmendorf and others 2012), has been a major component of the vegetation changes observed  
56 in the Arctic over the past few decades of global warming (Epstein and others 2013; Myers-  
57 Smith and others 2020). Rapid erect shrub expansion (c.f. shrubification) at the expense of non-  
58 vascular and herbaceous vegetation has been initially expected to increase soil organic carbon  
59 (SOC) stocks (Myers-Smith and others 2011), as plant carbon supply to soils is proportional to  
60 primary productivity (Metcalf and others 2018; Chen and others 2021), and decomposition of  
61 shrub leaf litter is slower than that of forbs and grasses (Cornelissen and others 2007). Models  
62 developed to estimate variation in ecosystem carbon balance with vegetation changes in  
63 northern latitudes have notably pointed towards this scenario (Qian and others 2010; Mekonnen  
64 and others 2018). Yet, greater erect shrub abundance has not necessarily increased soil organic  
65 carbon stocks (Parker and others 2021), which in various sites have even been lower beneath  
66 shrub-dominated communities than beneath adjacent heaths, meadows or tussock tundra  
67 (Wilmking and others 2006; Sørensen and others 2018). A better understanding of the common

68 mechanisms driving the fate of assimilated carbon in the wake of Arctic vegetation changes is  
69 thus needed, especially to bolster estimation of future Arctic ecosystem carbon storage.

70 Changes in tundra carbon stocks and cycling with greater erect shrub abundance can first be  
71 associated with shifts in plant-regulated soil microclimate and microbial community  
72 (Wallenstein and others 2007; Myers-Smith and Hik 2013; Kemppinen and others 2021;  
73 relation 1 in Figure 1). Two plant-induced processes can also occur concurrently: a significant  
74 increase in fresh energy-rich carbon supply (referred hereafter as plant carbon supply) with  
75 increasing litter quality and quantity (DeMarco and others 2014; Liang and others 2018;  
76 relation 2 in Figure 1), and a loss of recalcitrant energy-poor soil organic matter (SOM) to the  
77 atmosphere following plant-enhanced mineralization (c.f. positive priming; Fontaine and others  
78 2011; relation 3 in Figure 1). Thus, the long-term plant regulation of the net SOC balance is  
79 linked to the maintenance of the carbon-nutrient stoichiometry required for optimal plant  
80 metabolism (Zhen 2009; Maire and others 2013; relation 4 in Figure 1).

81 Plant-enhanced mineralization (i.e. positive priming) is a key process to determine long-  
82 term sequestration of carbon (C) and nutrient (N) in SOM (c.f. ‘bank mechanism’; Fontaine  
83 and Barot 2005; Perveen and others 2014). This process is regulated by the availability of  
84 nutrients in soil solution, and ultimately by plant N uptake (Fontaine and Barot 2005; Perveen  
85 and others 2014). Under conditions where plant N demand is lower than soil N supply, soil  
86 microbes are prone to sequester N in SOM through microbial immobilization (Martel and Paul  
87 1974), and plant-enhanced mineralization is rather low. This eventually leads to new SOM  
88 humification and strong soil C gain (Loya and others 2002; Kallenbach and others 2016). The  
89 fate of carbon stored in the soil then depends on how plant-derived C is efficiently fragmented  
90 and/or incorporated into microbial products and stabilized on minerals (Cotrufo and others  
91 2015).

92 In contrast, under conditions where plant N demand overcomes soil N supply, plant litter  
93 and photosynthate-C allocation to the rhizosphere enhance respiration of C-limited microbial  
94 communities, which in turn promote the degradation of recalcitrant nutrient-rich SOM and the  
95 release of mineral nutrients that become available for plants (Dijkstra and others 2009, 2013;  
96 Huo and others 2017). SOM microbial mining is thus related to plant N demand (Perveen and  
97 others 2014; Henneron and others 2020b), which is closely linked to plant nutrient uptake (Kou  
98 and others 2022). This process can at times be so important that it exceeds the formation of  
99 new SOM through humification of fresh C (Fontaine and others 2004; Terrer and others 2021),  
100 leading to soil C loss. To date, studies on soil functioning in tundra ecosystems, where plant  
101 growth is generally limited by low nutrient availability (e.g. phosphorus and nitrogen in Elser  
102 and others 2007), have highlighted both microbial N-limitation on SOC decomposition rates  
103 (Sistla and others 2012) and SOM degradation of N-rich SOM in many sites (Keuper and others  
104 2020). However, the extent to which particular functional traits associated with plant  
105 physiology and/or size drive plant N demand and SOM microbial mining has received less  
106 attention. This currently limits our understanding of how plant-environment interactions shape  
107 tundra carbon cycling, especially in the context of Arctic shrubification.

108 Meanwhile, other investigations have indicated also that plant species identity (Lynch and  
109 others 2018; Street and others 2020; Gavazov and others 2022), therefore different N-  
110 acquisition strategies, are key drivers of tundra soil organic carbon dynamics (Hewitt and others  
111 2019; Clemmensen and others 2021). Rates of plant nutrient demand and carbon supply are  
112 tightly linked to the leaf economics spectrum, contrasting resource acquisitive with  
113 conservative species (Henneron and others 2020a, b). Acquisitive species characterized by high  
114 leaf N content and high metabolic activity are more prone to stimulate SOM mining, whereas  
115 conservative species with low leaf N and low metabolic activity are more likely to induce new  
116 SOM humification (Henneron and others 2020b; Clemmensen and others 2021). Across the

117 tundra biome, plant communities have been subjected to a strong turnover of species marked  
118 by the arrival of acquisitive species such as erect shrubs exhibiting high leaf nitrogen and high  
119 specific leaf area (Bjorkman and others 2018). This pattern has nonetheless been strongly  
120 associated with wetter habitats, as erect shrub expansion appears to be constrained by limited  
121 soil moisture availability (Myers-Smith and others 2015). Conservative species would thus be  
122 expected to dominate in drier habitats. However, tundra vegetation has also the potential to  
123 respond plastically to environmental changes (Bret-Harte and others 2001; Hudson and others  
124 2011), and acquisitive species might be able to colonize a wide range of environments. Within-  
125 species variation in organic carbon stored in shrub biomass across environments has not yet  
126 been comprehensively quantified (relations 7 & 8 in Figure 1). This hampers our understanding  
127 of the extent of shrub control over soil organic carbon stocks across environmental conditions  
128 and how it would evolve if growing conditions improve.

129 This study aimed at determining the extent to which erect shrub abundance and  
130 environmental changes modulate long-term soil organic carbon stock dynamics in Arctic tundra  
131 ecosystems. Our work centered around *Salix richardsonii* Hook. at the northern edge of its  
132 distribution range, in a High Arctic alluvial fan landscape where it is the only erect shrub  
133 species encountered. The influence of environmental changes was examined by contrasting  
134 zones with different hydrological regime and sediment input intensity, where *S. richardsonii*  
135 occurs in patches and is expected to display variation in traits associated with C supply and N  
136 demand. Specifically, we used an observational approach to evaluate how SOC stocks vary as  
137 a function of vegetation (i.e. presence/absence of *S. richardsonii*) and hydrological regime (i.e.  
138 active vs. abandoned channels).

## 139 **Materials and methods**

### 140 **Study area**

141 The study took place in the Qarlikturvik valley of Bylot Island, Nunavut, Canada (73°09'N,  
142 79°57'W; Figure 2a). At the altitude of 20 m a.s.l, the mean total annual precipitation is 220  
143 mm (snow and rain), while the mean annual temperature and mean temperature of the warmest  
144 month (July) are -15°C and 6.1°C, respectively (Gauthier and others 2013). This 122 km<sup>2</sup> valley  
145 connects C-79 and C-93 glaciers to the Navy Board Inlet sea via a proglacial river (Coulombe  
146 and others 2022). It is mainly characterized by a low-centered polygon landscape (Fortier and  
147 Allard 2004) that is composed of wetlands dominated by grasses and sedges, and mesic  
148 environments that harbour a greater species richness including prostrate shrubs like *Salix*  
149 *arctica*, *Salix reticulata* and *Salix herbacea* (Gauthier and others 2013; Perreault and others  
150 2016). The valley is bounded to the North and South by plateaus of ~400 m made of poorly-  
151 consolidated shale and sandstone (Miall and others 1980). After deglaciation, stream incision  
152 and gullying in the plateaus formed alluvial fans at the base of the valley walls (Fig. 2a; Blair  
153 and McPherson 2009). These alluvial fans are grossly conical in shape, with a narrow apex, an  
154 arcuate frontal section, and a convex cross-section. They correspond to aggradational  
155 sedimentary deposits gradually widening with time, and comprise surface channels which  
156 receive sediment and nutrients from upslope throughout the plant growing season, migrate  
157 laterally with sedimentation, and can become temporarily or permanently abandoned (Rioux  
158 2021).

### 159 **Study design**

160 Fieldwork was carried out on 4-6 July 2015 along two alluvial fans located at the foot of the  
161 valley southern plateau (Figure 2a), and that were randomly selected from the fifteen fans along  
162 which *Salix richardsonii* occurs. Observations along each fan were conducted in two zones  
163 with contrasting hydrological regime and sediment input intensity: (i) active channels, where  
164 water flows yearly and sediment input is greater, and (ii) abandoned (i.e. inactive) channels,  
165 where water doesn't flow anymore and vegetation colonizes the bed of these channels (Figure

166 2b). Along each channel, a paired sampling was carried out in two vegetation conditions, that  
167 were ca. 3-5 m apart and defined by the absence or dominance of *Salix richardsonii* (c.f.  
168 ‘outside/beneath *SR*’ in Figure 2c). Three pairs of quadrats were randomly chosen along each  
169 channel, leading to a total of 24 experimental units (i.e. 2 alluvial fans x 2 channels x 3 paired  
170 replicas). At these locations, the height of *S. richardsonii* individuals averaged  $35 \pm 2$  cm, with  
171 no difference in height between channels ( $F = 1.0$ ,  $P = 0.99$ ). A preliminary  
172 dendrochronological study of largest branches showed that these individuals are at least 100-  
173 year-old (Bisson 2016).

#### 174 **Vegetation characterization and woody plant trait measurements**

175 Species richness and abundance were determined in each randomly deployed pair of 0.7 m x  
176 0.7 m quadrats. Abundance of woody and graminoid species, lichens and mosses were  
177 quantified using Daubenmire cover abundance classes (Daubenmire 1959). Aboveground  
178 biomass of woody and graminoid species was also measured, within a 0.2 m x 0.2 m quadrat  
179 set up at the bottom left corner of each 0.7 m x 0.7 m quadrat. Leaves were separated from  
180 stems for each of the three woody species encountered in these sites, namely *Salix richardsonii*,  
181 *S. arctica* and *S. reticulata*. Biomass samples were oven-dried at 65°C for a minimum of 72  
182 hours and weighed after drying.

183 Prior to plant collection, a picture was taken at a height of 1.50 m above each 0.2 m x 0.2 m  
184 quadrat to determine the total number of leaves of each of the three *Salix* species. A subsample  
185 of 3-20 leaves per *Salix* species per quadrat was additionally separated and stored in a plastic  
186 bag at 4°C until leaf area (cm<sup>2</sup>) was estimated back at the camp facilities using WinFOLIA  
187 software (Regent Instruments Inc, Quebec City, QC, Canada). These leaf samples were also  
188 oven-dried at 65°C for a minimum of 72 hours and weighed after drying. The following leaf  
189 traits were measured per *Salix* species per quadrat: leaf size (cm<sup>2</sup> leaf), leaf dry mass (DM, g),  
190 and leaf composition in C, N, P, K, Ca, Mg,  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$ . Elementary chemical analyses (leaf



191 C, N, P, K, Ca, Mg content) were performed using inductively coupled plasma-atomic emission  
192 spectroscopy (Plasma Model 40, PerkinElmer, Waltham, MA, USA) following a Mehlich-3  
193 acid extraction method. Leaf  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  isotopic composition was determined using an  
194 elemental analyzer coupled with an isotope ratio mass spectrometer (EA-IRMS, Agilent  
195 technology, Santa Clara, CA, USA).

196 For each *Salix* species, specific leaf area (SLA,  $\text{cm}^2 \text{ leaf g}^{-1} \text{ DM}$ ) was calculated as the ratio  
197 of leaf area to leaf dry mass; leaf area index ( $\text{LAI}_w$ ,  $\text{m}^2 \text{ leaf m}^{-2} \text{ soil}$ ), as the ratio of total leaf  
198 area (i.e. leaf size x total number of leaves in the quadrat) to corresponding quadrat area; total  
199 leaf nutrient content ( $\text{TLN}_w$ ,  $\text{kg m}^{-2} \text{ soil}$ ), as the product of leaf nutrient content ( $\text{LNuC}_w$ ; i.e.  
200 the cumulative leaf content in N, P, K, Ca and Mg) and leaf biomass. Community-weighted  
201 trait means (that is, the mean trait value of *Salix* species weighted by the abundance of each  
202 species) were calculated per quadrat for each functional trait aforementioned.

### 203 **Soil sampling and analyses**

204 Within each quadrat, three soil cores were collected using a 15-cm long and 6-cm diameter  
205 corer. When present, the moss layer (0-5.8 cm deep) was immediately separated and stored in  
206 a paper bag. Moss biomass was oven-dried at  $65^\circ\text{C}$  for a minimum of 72 hours and weighed  
207 after drying. Soil cores were extracted down to the thaw front depth at the date of sampling.  
208 They systematically exceeded the root front and had a total depth (i.e. depth of the thawed  
209 portion of soil) ranging between 6.7 and 9.2 cm without moss layer. Upon soil core extraction,  
210 two soil horizons were distinguished visually using the Canadian System of Soil Classification,  
211 and immediately separated:  $A_1$ , which corresponded to the upper  $1.8 \pm 0.3$  cm (mean  $\pm$  SE),  
212 and contained the majority of fine roots; and  $A_2$ , corresponding to the next  $5.8 \pm 0.2$  cm, and  
213 that was weakly colonized by roots (Figure 1c). For each quadrat, the three soil core replicates  
214 were pooled per soil horizon, and stored in plastic bags at  $4^\circ\text{C}$ .

215 Back in the laboratory, fresh soil samples were weighed, and air-dried at 25°C. They were  
216 then sieved at 2 mm to separate roots from coarse organic material (> 2 mm), and fine earth (<  
217 2 mm,  $M_t$ ). Coarse organic material and fine earth were then weighed separately. Root biomass  
218 was oven-dried at 65°C for a minimum of 72 hours, and weighed after drying. A first aliquot  
219 of ~10 g of fine earth was oven-dried at 105°C to determine water content (in %) as follows:  
220  $((\text{fresh weight} - \text{dry weight}) / \text{dry weight}) * 100$ . A second aliquot of ~30 g of fine earth was oven-  
221 dried at 60°C until constant weight ( $M_{od}$ ), and the following parameters were measured: pH in  
222 soil water; particle-size distribution, using a laser particle-size analyser (Laser Particle Sizer  
223 Analysette 22 MicroTec Plus, Fritsch, Idar-Oberstein, Germany); soil organic C and N content,  
224 after decarbonation by HCl fumigation; soil  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  isotopic composition, using the same  
225 methodology than for leaf  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  (see above).

226 The remainder of fine earth (12.8-33.5 g of soil air-dried at 25°C, and < 2 mm) was  
227 successively wet-sieved for two minutes at 1 mm, 250  $\mu\text{m}$  and 63  $\mu\text{m}$  to differentiate the  
228 particulate organic matter compartments (POM) as follows: coarse organic matter (cPOM, 2  
229 mm to 1 mm), medium organic matter (mPOM, 1 mm to 250  $\mu\text{m}$ ) and fine organic matter  
230 (fPOM, 250  $\mu\text{m}$  to 63  $\mu\text{m}$ ), respectively. Each of the sieved fractions was dried in an oven at  
231 60°C until a constant weight was obtained, weighed with a precision of  $\pm 0.0001$  g ( $M_{ad}$   
232 compartments:  $M_{ad\_cPOM}$ ,  $M_{ad\_mPOM}$ ,  $M_{ad\_fPOM}$ ), and then analyzed to obtain total soil organic C  
233 and total N content, as well as soil  $^{13}\text{C}$  and  $^{15}\text{N}$  isotopic composition. The C content of the  
234 mineral-associated OM (MAOM; Lavallee and others 2020) was subsequently estimated as the  
235 difference between  $\text{SOC}_{\text{tot}}$  and the C contained in the three measured POM compartments.

236 Finally, composite samples of cPOM, mPOM and fPOM from the A<sub>2</sub> horizon were used for  
237 all sites (12 samples in total) to analyze radiocarbon ( $^{14}\text{C}$ ) composition at the A.E. Lalonde  
238 AMS laboratory (University of Ottawa, Canada), following protocols detailed in Crann and  
239 others (2017). The fraction modern carbon,  $F^{14}\text{C}$ , was calculated as the ratio of the sample

240  $^{14}\text{C}/^{12}\text{C}$  to the standard  $^{14}\text{C}/^{12}\text{C}$  ratio (here Ox-II) measured in the same data block, and the  
241 result was normalized to  $\delta^{13}\text{C}$  (PDB) following correction for spectrometer and preparation  
242 fractionation. The  $\text{D}^{14}\text{C}$  was calculated as  $(\text{F}^{14}\text{C}-1)*1000$ , and the  $\Delta^{14}\text{C}$  as  $(\text{F}^{14}\text{C}*e(1950-$   
243  $\text{y})/8267)-1)*1000$  (Crann and others 2017).

## 244 **Quantifying soil organic carbon stocks**

245 The volumetric carbon content  $C_{vol}$  ( $\text{gC m}^{-3}$  soil) of fine earth and POM compartments was first  
246 calculated per quadrat as follows:

$$247 \quad C_{vol} = \frac{M_t}{M_{ad}} \cdot M_{od} \cdot C \cdot \frac{1}{V} \quad \text{Eq. 1}$$

248 where  $M_{od}$  (g) refers to the weight of the oven-dried aliquot of fine earth (see above),  $C$  ( $\text{g g}^{-1}$   
249 soil) is the total organic carbon content of the corresponding soil sample, and  $V$  ( $\text{m}^{-3}$ ) is the  
250 soil bulk volume that was calculated from the diameter of the corer and the length of each of  
251 the three soil cores sampled per quadrat and pooled per soil horizon. The ratio  $M_t/M_{ad}$  was used  
252 to account for the weight of POM compartments ( $M_{ad}$ ) compared to the total air-dried weight  
253 of fine earth ( $M_t$ ). Soil bulk volume included coarse elements which contributed marginally to  
254 soil C content. Coarse elements were present in 64% of the soil samples, and when present  
255 accounted for only 2% of soil bulk volume, with no difference between vegetation conditions  
256 and channel zones ( $P > 0.33$  in both cases).

257 The surfacic (i.e. area based) carbon content  $C_{surf}$  ( $\text{gC m}^{-2}$  soil) of fine earth and POM  
258 compartments was calculated as follows:

$$259 \quad C_{surf} = \frac{M_t}{M_{ad}} \cdot M_{od} \cdot C \cdot \frac{1}{S} \quad \text{Eq. 2}$$

260 where  $S$  ( $\text{m}^2$ ) is the soil surface that was obtained from the diameter of the corer.

## 261 **Soil temperature monitoring**

262 Soil temperature was assessed with HOBO Pendant 64K data loggers (Onset Computer  
263 Corporation, Bourne, MA, USA; accuracy of  $\pm 0.53^{\circ}\text{C}$  from  $0^{\circ}$  to  $50^{\circ}\text{C}$ , resolution of  $0.14^{\circ}\text{C}$   
264 at  $25^{\circ}\text{C}$ ), that were deployed in three paired vegetation conditions (outside and beneath *S.*  
265 *richardsonii*) along one of the two alluvial fans studied (1 fan x 2 channels x 3 paired replicas  
266 = 12 loggers). Each logger recorded soil temperature at 6 cm depth hourly during one year  
267 between July 2017 and July 2018. Freezing degree-days (FDD) were calculated as the  
268 cumulative sum of daily average negative temperatures between summer 2017 and summer  
269 2018, while thawing degree-days (TDD) were calculated as the cumulative sum of daily  
270 average positive temperatures for the years 2017 and 2018.

## 271 **Statistical analyses**

272 *Testing the effect of hydrological regime and Salix richardsonii occurrence on plant*  
273 *communities and soil conditions* - We first ran linear mixed models (MIXED procedure with  
274 REML method in SAS 9.4, SAS Institute, Cary, NC) to explain variation in plant traits, soil  
275 conditions and SOC with changes in hydrological regime and occurrence of *S. richardsonii* .  
276 Channel (active vs abandoned), vegetation condition (outside vs beneath *S. richardsonii*), and  
277 the interaction between channel and vegetation condition were treated as fixed factors, with  
278 random effects structured as pairs of quadrats nested within channels within fans. We checked  
279 prior to analyses that the random effects were normally distributed. Evaluation of significant  
280 effects was based on an alpha of 0.05. As multiple comparisons were performed, the  
281 significance of results was further assessed using the False Discovery Rate procedure  
282 (Benjamini and Hochberg 1995; but see Pike 2011 for the method applied in ecology and  
283 evolution). In addition, we quantified (i) the association between plant community and soil  
284 conditions with a multivariate canonical correspondence analysis (CCA; ter Braak and  
285 Verdonschot 1995), and (ii) the association between channel, vegetation conditions, and leaf

286 nutrients of *Salix* spp. with a principal component analysis (PCA), using the *vegan* package in  
287 R version 4.0.4 (R Development Core Team 2021).

288 *Testing the influence of plant carbon supply and nutrient demand on soil organic carbon stocks*

289 - This analysis was conducted using a two-step approach. First, we selected the measured  
290 community-weighted traits (i.e. variables) that were associated with C supply and N demand,  
291 and could best represent the influence of the two processes on soil organic carbon stock  
292 variation. Specifically, we used (i)  $SLA_w$ ,  $LAI_w$ , and moss biomass as proxies of plant C supply,  
293 considering in particular that  $SLA_w$  and  $LAI_w$  are associated with plant fast resource acquisition  
294 and rapid growth (Wright and others 2004; Sterck and others 2011), and with gross and net  
295 primary productivity (Shaver and others 2013); (ii) leaf nutrient content ( $LNuC_w$ ),  
296 leaf/stem/root biomass, and  $TLN_w$  as proxies of plant nutrient demand, considering that the  
297 biomass of leaves, stems and roots is directly related to the cost of tissue maintenance that  
298 requires soil nutrients (Givnish and others 2014). A multi-model inference that included the  
299 eight aforementioned explicative variables was performed to investigate the sources of  
300 variation in soil organic carbon stocks. We used the *MuMIn* package in R, and a linear mixed  
301 regression model with random effects structured as pairs of quadrats nested within channels  
302 within fans. We used model weights to estimate the relative importance of each variable under  
303 consideration (*dredge* function). We checked for model convergence (*nloptwrap* function), and  
304 used AICc as the ranking-based index. Then, we selected the most important variable for each  
305 of the carbon supply and nutrient demand process (*importance* function). The two selected  
306 variables were included in a path analysis that was performed based on the series of mixed  
307 regressions identified in Figure 1, using the *piecewiseSEM* package in R (Lefcheck 2016).  
308 Variation in SOC stocks was modelled with multiple linear mixed regression models ( $n = 24$   
309 experimental units; *nlme* package in R), using plant carbon supply, plant nutrient demand,  
310 hydrological regime and occurrence of *S. richardsonii* as fixed factors. Pairs of quadrats nested

311 within channels within fans were considered as random factors. Implied directed separations  
312 were tested, and associated *P*-values used to compute the model goodness-of-fit.

313 *Testing the influence of plant carbon supply and nutrient demand on soil organic carbon*  
314 *quality* - Signatures of soil  $\delta^{13}\text{C}$ ,  $^{14}\text{C}$  and  $\delta^{15}\text{N}$  were used to estimate the change in SOC quality  
315 that has resulted from variation in soil carbon gain and OM degradation associated with  
316 vegetation change (i.e. occurrence of *S. richardsonii*). Given that SOC stocks in the Arctic  
317 mainly come from large plant residues with relatively little microbial processing (see e.g. Prater  
318 and others 2020), differences in soil bulk  $\delta^{13}\text{C}$  observed between vegetation conditions were  
319 expected to result from the input of newly formed plant litter that is depleted in  $\delta^{13}\text{C}$  as a  
320 consequence of the Suess  $\text{CO}_2$  fertilization effect (Ehleringer and others 2000; Camino-Serrano  
321 and others 2019). Thus, the abundance of *S. richardsonii* and related additional carbon supply  
322 were assumed to decrease soil bulk  $\delta^{13}\text{C}$  (i.e. leading to more negative values). We used soil  
323 bulk  $^{14}\text{C}$  to visually test how vegetation conditions modified the average age of SOC, with more  
324 positive values indicating more recent SOC.

325 Conversely to carbon isotopes, there is a significant microbial discrimination against  $^{15}\text{N}$   
326 during OM mineralization when microbial growth is C-limited (Dijkstra and others 2009). This  
327 leads to excretion of depleted mineral N and accumulation of  $\delta^{15}\text{N}$  in microbial biomass, which  
328 ultimately increases soil bulk  $\delta^{15}\text{N}$  values (Dijkstra 2009; Craine and others 2015).  
329 Theoretically, enrichment of soil  $\delta^{15}\text{N}$  might also originate from N losses through volatilization,  
330 denitrification and N leaching; however, the effect of these N losses on soil  $\delta^{15}\text{N}$  signature is  
331 likely marginal as the substantial increase in plant biomass related to the occurrence of *S.*  
332 *richardsonii* is expected to reduce these N losses. Soil bulk  $\delta^{15}\text{N}$  values might also be modified  
333 by (i) N fixation - in this case, while N-fixing leguminous species such as *Astragalus alpinus*  
334 were rare in our sampling plots,  $\delta^{15}\text{N}$  might tend to zero due to N fixation by diazotrophs present  
335 on mosses (Rousk and others 2017); and (ii) arbuscular and ectomycorrhizal associations

336 established by *Salix* spp. (Dhillion 1994) - in this case, there may be a  $^{15}\text{N}$  depletion in plants  
337 and  $^{15}\text{N}$  enrichment in mycorrhizal fungi (Hobbie and Hobbie 2009), likely leading to a zero-  
338 sum for bulk soil  $\delta^{15}\text{N}$ .

339 The effects of plant carbon supply and nutrient demand on soil bulk  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  across  
340 soil horizons and OM compartments were tested with generalized linear mixed models, where  
341 soil horizons and OM compartments as well as their interaction terms with plant carbon supply  
342 and nutrient demand were treated as fixed factors, and pairs of quadrats as well as fans nested  
343 within channel as random factors.

## 344 **Results**

### 345 **Variation in plant communities**

346 There was no significant difference between active and abandoned channels in *S. richardsonii*  
347 abundance (non-significant channel effect;  $P = 0.08$ ), plant species richness ( $P = 0.72$ ), and  
348 plant community (Supplementary materials 1-3).

349 As expected, abundance of *S. richardsonii* was significantly greater in plots sampled beneath  
350 (> 50% cover) than in plots sampled outside (< 0.5%) the species' patches (significant  
351 vegetation effect;  $P < 0.001$ ; Supplementary material 1). The difference in *S. richardsonii*  
352 abundance between vegetation conditions did not translate into a difference in plant species  
353 richness, which remained low (5-7 species) both beneath and outside the species' patches ( $P =$   
354  $0.08$ ; Supplementary material 1). However, we observed a change in plant community  
355 composition, with living mosses and vascular plant species such as *Astragalus alpinus*,  
356 *Arctagrostis latifolia*, *Dryas integrifolia*, *Salix artica* and *S. reticulata* occurring in plots  
357 located outside *S. richardsonii* patches (Supplementary material 3).

358 The CCA testing the association between plant community and soil conditions further  
359 showed that soil variables (i.e. thaw front depth, bulk density, water content, pH, FDD, TDD,

360 C:N ratio, and total N content), hydrological regime and vegetation conditions together  
361 explained 57% of the total variation in species abundance, with the first two axes retaining 32%  
362 and 9% of the total variation, respectively (Supplementary material 4).

### 363 **Variation in plant traits**

364 No differences in plant biomass-related traits were found between active and abandoned  
365 channels (non-significant channel effect;  $P > 0.07$  in all cases), with the exception of moss  
366 biomass that was significantly greater in abandoned channel zones (Table 1).

367 Regardless of the hydrological regime, significant differences in plant traits were observed  
368 between vegetation conditions, with the exception of root and moss biomass (Table 1,  
369 Supplementary material 1). For instance, plant aboveground biomass ranged from 112 gDW  
370  $\text{m}^{-2}$  soil in plots outside *Salix richardsonii* to 1416 gDW  $\text{m}^{-2}$  soil beneath the species' patches  
371 (significant vegetation effect;  $P < 0.001$ ; Table 1, Supplementary material 5), and was driven  
372 by a 10 to 30 times greater aboveground biomass of *S. richardsonii* in plots beneath the species'  
373 patches ( $P < 0.001$ ; Table 1). Aboveground biomass of graminoids was 2 to 5 times greater in  
374 plots outside *S. richardsonii* patches. Stem and leaf biomass of *S. richardsonii* were  
375 respectively 95% and 70% greater than that of *Salix arctica* and *S. reticulata* ( $P < 0.001$  and  $P$   
376  $= 0.009$ , respectively; Table 1). *S. richardsonii* exhibited significantly more leaves, larger  
377 leaves, and lower SLA than the other two *Salix* species ( $P < 0.001$  in all cases; Supplementary  
378 material 1). Thus,  $\text{LAI}_w$  and  $\text{TLN}_w$  beneath *Salix richardsonii* were respectively twice and three  
379 times as big as in plots outside *S. richardsonii* patches ( $P < 0.001$  and  $P = 0.006$ , respectively;  
380 Table 1).

381 The interaction between hydrological regime and occurrence of *S. richardsonii* led to  
382 significant differences in  $\text{LAI}_w$  ( $r^2 = 0.73$ ,  $P < 0.01$  for all additive and interactive terms). The  
383 difference was particularly strong between vegetation conditions along active channels, where  
384  $\text{LAI}_w$  values were very low in plots outside *S. richardsonii* patches (Table 1).



385 The PCA testing the association between channel, vegetation conditions and leaf nutrients  
386 of *Salix* spp. showed that *Salix* leaves also discriminated strongly against plots differing in the  
387 *S. richardsonii* occurrence (Table 1, Supplementary materials 1 and 4). For instance, leaves of  
388 *S. richardsonii* exhibited significantly 4% lower C, 14% higher N, 16% lower C:N ratio, and  
389 16% higher nutrient content (LNuC<sub>w</sub>) compared to leaves of other *Salix* species found in plots  
390 outside *S. richardsonii* patches (Supplementary material 1). No difference in leaf biochemistry  
391 was observed between active and abandoned channels ( $P > 0.16$  in all cases; Table 1,  
392 Supplementary material 1).

### 393 **Variation in soil conditions**

394 At the time of sampling in early summer, some differences in soil conditions arose between  
395 hydrological regimes, i.e. between active and abandoned channels. While thaw front depth,  
396 depth of each soil horizon, and soil bulk density were similar between hydrological regimes,  
397 soils along active channels exhibited significantly 6% higher pH and 30% lower total water  
398 content than those sampled along abandoned channels (Table 1, Supplementary material 5).  
399 Active and abandoned channels also differed significantly in soil temperature ( $P = 0.001$  and  
400  $P = 0.02$  for FDD and TDD, respectively; Table 1), with abandoned channels showing 26%  
401 higher FDD and 44% lower TDD than active channels (Supplementary material 5). No  
402 difference in soil C:N ratio and soil total N content emerged between channel zones ( $P = 0.15$   
403 and  $P = 0.64$ , respectively; Table 1).

404 Patches of *S. richardsonii* along active and abandoned channels were not associated with  
405 changes in thaw front depth and soil bulk density compared to plots dominated by prostrate  
406 shrubs and graminoids (non-significant vegetation effect,  $P > 0.19$  in both cases; Table 1). Soil  
407 total water content and pH measured beneath *S. richardsonii* averaged 3.9% and 7.4,  
408 respectively, i.e. within the range of values measured in plots outside *S. richardsonii* patches  
409 ( $P > 0.43$  in both cases; Supplementary material 5). Patches of *S. richardsonii* were not

410 associated either with variation in soil freezing and thawing degrees-days measured across two  
411 seasons ( $P > 0.26$  in both cases), which respectively averaged  $5700^{\circ}\text{C}$  and  $240^{\circ}\text{C}$  in both  
412 vegetation conditions (Table 1, Supplementary material 5). There was no difference in soil C:N  
413 ratio and soil total N content between vegetation conditions ( $P = 0.33$  and  $P = 0.28$ ,  
414 respectively; Table 1). Yet, along active channels soil total N content increased by 37% beneath  
415 *S. richardsonii* compared to plots dominated by prostrate shrubs and graminoids ( $P = 0.05$ ).

#### 416 **Variation in total soil organic carbon stocks**

417 Total soil organic carbon stocks ( $\text{SOC}_{\text{tot}}$ ) averaged  $18.0 \pm 0.6 \text{ kgC m}^{-3}$  (equivalent to  $1.4 \pm 0.1$   
418  $\text{kgC m}^{-2}$ ) across all plots (volumetric SOC data are presented in the main text; see  
419 Supplementary materials 1 and 6 for surfacic SOC data). There was no significant difference  
420 in  $\text{SOC}_{\text{tot}}$  between hydrological regimes, nor between vegetation conditions. While no  
421 difference in  $\text{SOC}_{\text{tot}}$  arose between vegetation conditions located along abandoned channels ( $P$   
422  $> 0.60$ ),  $\text{SOC}_{\text{tot}}$  differed between vegetation conditions along active channels, with a 18-28%  
423 increase in organic C in soils sampled beneath *S. richardsonii* compared to those sampled  
424 outside the species' patches ( $r^2 = 0.15$  and  $P = 0.08$  for volumetric content,  $r^2 = 0.46$  and  $P =$   
425  $0.02$  for surfacic content; Figure 3a, Supplementary material 6).

#### 426 **Drivers of soil organic carbon stocks**

427 The model selection indicated that  $\text{LAI}_w$  and  $\text{TLN}_w$  were the variables that best represented the  
428 influence of plant C supply and N demand on  $\text{SOC}_{\text{tot}}$  variation, respectively (Figure 4a). The  
429 path analysis revealed that  $\text{SOC}_{\text{tot}}$  was related positively to  $\text{LAI}_w$  and negatively to  $\text{TLN}_w$  ( $\rho =$   
430  $0.86$ ,  $P < 0.01$  and  $\rho = -0.64$ ,  $P < 0.05$ , respectively; Figure 4b,c), and that together these  
431 relations increased the amount of  $\text{SOC}_{\text{tot}}$  variation explained ( $r^2 = 0.47$ ; Table 2, Figure 4d),  
432 when compared with the categorical model ( $r^2 = 0.15$ , see earlier paragraph). The path analysis  
433 also retained the covariation between  $\text{LAI}_w$  and  $\text{TLN}_w$  ( $P = 0.054$ ), the positive effect of *S.*

434 *richardsonii* occurrence on LAI<sub>w</sub> and TLN<sub>w</sub> ( $\rho = 0.79$ ,  $P < 0.001$ , and  $\rho = 0.55$ ,  $P < 0.01$ ,  
435 respectively), as well as the positive effect of hydrological regime on LAI<sub>w</sub> and TLN<sub>w</sub>, although  
436 the latter two relations were not significant ( $\rho = 0.12$  and  $\rho = 0.40$ , respectively; Figure 4d). In  
437 contrast, the path analysis did not retain the direct effects of hydrological regime and *S.*  
438 *richardsonii* occurrence on SOC<sub>tot</sub>, indicating that there was no residual effect of vegetation  
439 and environmental changes on total soil organic carbon stock.

#### 440 **Variation in soil organic carbon stocks across soil horizons and OM compartments**

441 A significant difference in soil organic carbon stocks was observed between soil horizons, with  
442 stocks being consistently higher in the top soil horizon A<sub>1</sub> than in the soil horizon A<sub>2</sub> (30.0 vs  
443 15.2 kgC m<sup>-3</sup>, respectively;  $P < 0.001$ ; Table 1, Figure 3b). There was no difference in SOC-  
444 A<sub>1</sub> and SOC-A<sub>2</sub> between hydrological regimes ( $P > 0.40$  in both cases), nor between vegetation  
445 conditions (Table 1). SOC contained in horizons A<sub>1</sub> and A<sub>2</sub> did not differ between vegetation  
446 conditions along abandoned channels ( $P = 0.27$  in the two cases; Table 1, Figure 3b). Yet,  
447 differences arose between vegetation conditions along active channels, with SOC-A<sub>1</sub> beneath  
448 *S. richardsonii* decreasing significantly by 21% compared to plots located outside the species'  
449 patches (27.3 vs 34.6 kgC m<sup>-3</sup>;  $P = 0.04$ ; Table 1, Figure 3b). SOC-A<sub>2</sub> also showed a 25%  
450 increase beneath *S. richardsonii* (16.7 vs 12.4 kgC m<sup>-3</sup>), but the difference between vegetation  
451 conditions was not statistically significant ( $P = 0.14$ ; Table 1, Figure 3b).

452 Analyses of SOC per OM compartment showed that soil organic carbon was consistently  
453 higher in OM compartments beneath *S. richardsonii*, the difference between vegetation  
454 conditions being particularly marked for mPOM and fPOM compartments (Table 1,  
455 Supplementary material 1). This pattern was driven by significant differences in C-mPOM and  
456 C-fPOM of the soil horizon A<sub>2</sub> along active channels, where both C-mPOM and C-fPOM were  
457 > 85% higher beneath than outside *S. richardsonii* patches ( $P = 0.03$  in both cases; Table 1,  
458 Figure 3c). Contrastingly, no significant difference between vegetation conditions was found

459 in C-cPOM regardless of the hydrological regime and soil horizon, nor in C-mPOM and C-  
460 fPOM along abandoned channels ( $P > 0.21$  in all cases; Figure 3c). The C content of the  
461 mineral-associated OM (MAOM) did not exhibit overall differences between vegetation  
462 conditions and channels ( $P > 0.12$  in all cases). Yet, C-MAOM of the soil horizon A<sub>1</sub> was lower  
463 beneath *S. richardsonii* along active channels ( $P = 0.05$ ; Figure 3c).

#### 464 **Quality of soil organic carbon**

465 Across all plots, the increase in soil organic carbon associated with greater plant carbon supply  
466 was characterized by a change in the OM <sup>13</sup>C and <sup>14</sup>C isotopic signature. Soil bulk  $\delta^{13}\text{C}$  was  
467 strongly and negatively associated with LAI<sub>w</sub> ( $r^2 = 0.58$ ,  $P < 0.001$ ; Figure 5a). This effect was  
468 particularly noticeable in the horizon A<sub>1</sub> compared to A<sub>2</sub> ( $r^2 = 0.50$ ,  $P < 0.001$  and  $r^2 = 0.33$ ,  $P$   
469  $= 0.001$  for A<sub>1</sub> and A<sub>2</sub>, respectively; Figure 5b), and in each OM compartment ( $r^2 > 0.35$  and  $P$   
470  $< 0.001$  in the three cases; Figure 5c). The lowest soil  $\delta^{13}\text{C}$  values were found under high net  
471 carbon supply conditions, i.e. in plots sampled beneath *S. richardsonii* regardless of the  
472 hydrological regime, and were closer to leaf  $\delta^{13}\text{C}$  signature compared to  $\delta^{13}\text{C}$  of soils sampled  
473 outside *S. richardsonii* patches (Table 1, Supplementary material 7). Variation in SOM isotopes  
474 with vegetation change was also captured when analyzing OM  $\Delta^{14}\text{C}$  values of soil horizon A<sub>2</sub>.  
475 Values of  $\Delta^{14}\text{C}$ , which were lower in active than in abandoned channels for each POM  
476 compartment considered, were always higher (i.e. less negative) in plots beneath *S. richardsonii*  
477 than in plots dominated by herbaceous and other *Salix* spp. (Figure 6). The difference in  $\Delta^{14}\text{C}$   
478 between vegetation conditions was particularly marked when considering the A<sub>2</sub>-fPOM  
479 compartment sampled along active channels (94.26‰). By contrast, the difference in the A<sub>2</sub>-  
480 fPOM  $\Delta^{14}\text{C}$  between vegetation conditions along abandoned channels was of 14.88‰.

481 Plant N demand was positively but weakly related to soil bulk  $\delta^{15}\text{N}$  ( $r^2 = 0.13$ ,  $P = 0.07$ ;  
482 Figure 5d). The same pattern was observed in the soil horizon A<sub>2</sub> ( $r^2 = 0.11$ ,  $P = 0.09$ ; Figure  
483 5e) and in the A<sub>2</sub>-cPOM compartment ( $r^2 = 0.09$ ,  $P = 0.15$ ; Figure 5f). Soil  $\delta^{15}\text{N}$  values were

484 generally less negative than the leaf  $\delta^{15}\text{N}$  signature (Table 1, Supplementary material 7). The  
485 highest (i.e. less negative) soil  $\delta^{15}\text{N}$  values were found under high nutrient demand, i.e. in plots  
486 beneath *S. richardsonii* regardless of the hydrological regime (Figure 5d).

## 487 **Discussion**

488 This study aimed at exploring how tundra shrubification and environmental changes affect soil  
489 organic carbon stock dynamics in the High Arctic. We used the occurrence of *Salix richardsonii*  
490 patches along channels differing in water and nutrient availability to investigate the extent to  
491 which changes in soil organic carbon stocks were associated with a shift in plant economic  
492 strategies. Our observational approach suggested that increasing plant productivity associated  
493 with greater erect shrub abundance can lead to an increase in soil organic carbon stock  
494 compared to adjacent plant communities dominated by prostrate shrubs and graminoids.  
495 However, the greater soil organic carbon stocks beneath *S. richardsonii* patches were not linked  
496 to direct effects of plant taxa composition or environmental changes (relations 1 and 9 in Figure  
497 1), but rather to variation in plant aboveground functional traits associated with greater carbon  
498 supply and nutrient demand (relations 2 and 3 in Figure 1).

### 499 ***S. richardsonii* increased total soil organic carbon stocks along active channels of alluvial** 500 **fans**

501 We found that total soil organic carbon stocks beneath *S. richardsonii* patches ranged from 1.05  
502 to 2.00 kg m<sup>-2</sup> (14.76 to 22.68 kg m<sup>-3</sup>). Conducted at a latitude of 73°N in comparison with the  
503 62°-68°N latitudinal range of previous work (e.g. Wilmking and others 2006; Hartley and  
504 others 2012; Sørensen and others 2018; Street and others 2018; Parker and others 2021), this  
505 study represents so far the northernmost evaluation of soil organic carbon stocks in the context  
506 of increasing erect deciduous shrub abundance in tundra ecosystems. The stocks we found are  
507 within the range of soil organic carbon pools that have been reported to date in the handful of

508 other erect shrub-dominated tundra sites, going from 0.93 kg m<sup>-2</sup> under *Salix glauca* L. and  
509 *Salix lapponum* L. in Dovre Mountain alpine tundra, Norway (Sørensen and others 2018) and  
510 1.66-2.79 kg m<sup>-2</sup> under *Betula nana* L. and *Salix pulchra* Cham. in low Arctic tundra near  
511 Toolik Lake, AK, USA (DeMarco and others 2011; Lynch and others 2018) to 3.00 ± 0.50 kg  
512 m<sup>-2</sup> under *Betula nana* L. and *Salix* spp. thickets in low Arctic tundra near Abisko, Sweden  
513 (Parker and others 2015; Parker and others 2020), 3.64 ± 1.40 kg m<sup>-2</sup> under *Alnus viridis*  
514 (Chaix) DC. in low Arctic tundra near Inuvik, NWT, Canada (Street and others 2018) and 6.00  
515 kg m<sup>-2</sup> under *Betula glandulosa* Michx. in subarctic tundra near Umiujaq, QC, Canada (Gagnon  
516 and others 2019).

517 Our observations underscore the potential of *S. richardsonii* to increase soil organic carbon  
518 stocks compared to neighboring prostrate tundra when local environmental conditions are  
519 suitable. This plant-environment interaction observed from a fine-scale paired sampling is  
520 particularly striking considering that changes in soil organic carbon stocks are usually difficult  
521 to detect due to their large pool size, slow turnover rate and substantial spatial heterogeneity  
522 (van Groenigen and others 2014). As *S. richardsonii* in the studied sites are probably older than  
523 100 years (Bisson 2016), further analyses are now required using root collars, which might be  
524 buried deep considering the sedimentation process occurring in this system, to determine the  
525 period of time over which such shrub-mediated increase in soil organic carbon stocks has  
526 occurred. In addition, a recent evaluation of soil carbon stocks of dominant geomorphological  
527 terrain units conducted in the same valley as our study sites showed that alluvial fans can store  
528 up to 25 kgC m<sup>-3</sup> within the first 5 cm of soil (Ola and others 2022), which is in the same order  
529 of magnitude as the SOC values we found. It now remains to be seen how variation in  
530 catchment size, channel slope, run off, and sedimentation rates among alluvial fans of the valley  
531 influence these findings.

532 The increase in soil organic carbon stocks beneath *S. richardsonii* along active channels of  
533 alluvial fans differs from the majority of carbon cycle studies that were carried out among low  
534 arctic and subarctic tundra ecotones, and that reported either no change (Lynch and others 2018)  
535 or a depletion in soil organic carbon pools in erect shrub sites compared to adjacent heath and  
536 meadow systems (e.g. Wilmking and others 2006; Hartley and others 2012; Sørensen and  
537 others 2018). The discrepancy among studies may be in part explained by the fact that they  
538 primarily linked soil carbon storage to changes in plant community composition, regardless of  
539 the functional mechanisms of soil carbon stock variation. This is also illustrated by the results  
540 of Kemppinen and others (2021), who found that soil organic carbon stocks were positively  
541 related to overall plant coverage and woody plant height, and negatively to woody plant  
542 dominance. Accounting only for changes in plant taxa composition or plant functional groups  
543 might thus limit our mechanistic understanding of the effects of increasing erect shrub  
544 abundance on soil carbon storage. Alternatively, we found that the magnitude of variation in  
545 soil organic carbon stocks can be affected by the depth of soil horizons considered. While most  
546 studies have to date reported soil carbon stocks per unit surface (Parker and others 2021), also  
547 providing estimates of soil carbon stocks per unit volume might further inform about how  
548 organic carbon is distributed along soil profiles.

#### 549 **Biological drivers of soil organic carbon stock variation**

550 Soil conditions such as total water content, thermal status and thaw front depth were similar  
551 beneath and outside *S. richardsonii* patches, and are thus unlikely to explain the contrasting  
552 soil organic carbon stocks between vegetation conditions (relation 1 in Figure 1). In the active  
553 channel zones characterized by regular water flows and nutrient inputs, the higher carbon stocks  
554 beneath *S. richardsonii* may rather be a consequence of the response of plant resource  
555 acquisition traits to favorable growing conditions. Indeed, *S. richardsonii* exhibited higher LAI  
556 and greater leaf and stem biomass compared to adjacent vegetation dominated by prostrate

557 *Salix arctica* and *S. reticulata*, leading to higher plant carbon supply that outweighed potential  
558 carbon losses from soil respiration (relation 5 in Figure 1). Comparatively, the increase in soil  
559 organic carbon stocks beneath *S. richardsonii* patches did not hold true in the abandoned  
560 channel zones. In these habitats with more limited water flows and nutrient inputs, the reduction  
561 in LAI compared to active channel zones may be responsible for the lack of influence of *S.*  
562 *richardsonii* dominance on soil organic carbon stocks. Such trait variation along abiotic  
563 gradients supports observations throughout the tundra that plant trait attributes strongly respond  
564 to local variability in environmental conditions (Bret-Harte and others 2001; Bjorkman and  
565 others 2018).

566 Irrespective of channel zones and vegetation conditions, soil organic carbon decreased for a  
567 given LAI with increasing TLN (relation 3 in Figure 1). This suggests that increasing nutrient  
568 demand has likely led to a N-limiting situation, which has stimulated plant-enhanced  
569 mineralization, in turn inducing a decrease in SOC. In line with all other studies showing that  
570 tundra is particularly limited by nutrients (Chapin and others 1995; Elser and others 2007;  
571 Keuper et al. 2020), our result highlighted the importance of considering plant nutrient demand  
572 when quantifying the influence of vegetation changes and plant-enhanced mineralization on  
573 soil carbon content. They support observations by Henneron and others (2020b), who found  
574 that differences in soil organic carbon mineralization among temperate grassland species were  
575 related to interspecific variation in economic strategies that sustain nutrition. Specifically, soil  
576 under fast-growing species with higher photosynthetic rates and greater aboveground biomass  
577 exhibited higher carbon mineralization than soil under slow-growing conservative species  
578 (Henneron and others 2020b). While photosynthetic rates have yet to be measured at our study  
579 sites, we still found that soil carbon content beneath *S. richardsonii* was associated with high  
580 leaf biomass and related high total nutrient demand. This further indicates that biomass-related  
581 traits such as LAI are key features to consider when studying how plant nutrition controls soil



582 C dynamics, in addition to the traits captured within the leaf economics spectrum (LES;  
583 Henneron and others 2020a; Terrer and others 2021). For instance, leaf N content, which  
584 belongs to the LES, does not necessarily correlate with plant N demand, which belongs to the  
585 plant height-biomass spectrum (Maire and others 2009).

586 According to our assumption, we showed that a functional trait-based approach is beneficial  
587 to provide new insights into how vegetation changes regulate tundra carbon cycling. Through  
588 the path analysis, we showed that only traits were required to explain SOC variation in the  
589 studied system, whereas all the other vegetation conditions such as moss and root biomass were  
590 not. Our study agrees with the findings of Happonen and others (2022), who showed that  
591 aboveground plant traits had strong relationships with either aboveground or soil carbon stocks.  
592 It also highlights the relevance of using LAI to characterize vegetation effects on carbon cycling  
593 in the tundra (Shaver and others 2007; Street and others 2007; Parker and others 2020). The  
594 fact that in the study of Happonen and others (2022) soil carbon stocks were significantly  
595 related to SLA and LDMC instead of plant height further suggests that measuring leaf economic  
596 traits, that integrate plant investment in carbon and nutrients (Wright and others 2004), are key  
597 for a better mechanistic understanding of vegetation-carbon interactions. The strong and  
598 negative relationship between soil organic carbon stocks and TLN demonstrates that  
599 considering functional traits related to plant nutrient demand in parallel of those associated with  
600 carbon supply is important to better characterize plant-soil linkages that shape the fate of soil  
601 carbon stocks in tundra landscapes.

602 The explanatory power of our models was relatively high even if we only considered a few  
603 plant aboveground traits as proxies of carbon supply and nutrient demand dynamics.  
604 Particularly, all the other vegetation conditions not associated with traits were not required in  
605 our path analysis model (relation 1 in Figure 1). Yet, vegetation changes also induce significant  
606 shifts in plant-mycorrhizal associations and microbial community (Deslippe and others 2011;

607 Clemmensen and others 2021). Shifts in soil carbon storage between vegetation conditions are  
608 likely to be also driven by rhizosphere microbiome and fine rooting traits involved in plant-soil  
609 feedbacks (Parker and others 2021; Spitzer and others 2022). Further attention on belowground  
610 processes will thus be required, especially in sites where increasing erect shrub abundance is  
611 associated with changes in woody plant taxa.

## 612 **Regulation of soil isotopic signatures**

613 Differences between vegetation conditions in soil stable  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  and radioactive  $^{14}\text{C}$   
614 isotopes illustrated that *S. richardsonii* has strongly influenced soil chemistry and stability of  
615 SOM stocks, in accordance with the well documented effects of plant economic traits on soil  
616 biogeochemistry (Zhu and others 2016; Henneron and others 2020a). These results also align  
617 with changes in soil carbon supply and nutrient demand dynamics. Greater  $\text{LAI}_w$  (i.e. greater  
618 carbon supply) was associated with more negative  $\delta^{13}\text{C}$  in all soil horizons and OM  
619 compartments (Supplementary material 7), suggesting that new plant-derived carbon, depleted  
620 in  $^{13}\text{C}$  because of the Suess effect (Ehleringer and others 2000), has been incorporated into all  
621 soil compartments (Figure 5, Supplementary material 7). The steeper slope of the  $\delta^{13}\text{C}$ - $\text{LAI}_w$   
622 relationship for the top soil horizon  $A_1$  compared to the soil horizon  $A_2$  (Figure 5b) indicates  
623 that the input of new carbon may be greater in the first two centimeters of soils, where we found  
624 the vast majority of roots. However, the amount of new carbon in the soil horizon  $A_2$  along  
625 active channels was significant enough to increase soil organic carbon stocks and rejuvenate  
626 the  $\Delta^{14}\text{C}$  signature of the fPOM compartment from  $-291\text{‰}$  to  $-197\text{‰}$  (Figure 6), which is  
627 equivalent to a mean age shift from  $2809 \pm 86$  to  $1619 \pm 80$  years (based on Crann and others  
628 2017). We also observed a higher elevation of the  $\delta^{13}\text{C}$ - $\text{LAI}_w$  relationship for the fPOM  
629 compartment (Figure 5c). It is likely that the fPOM compartment incorporated minerals  
630 associated with less-depleted carbon compounds such as carbohydrate, sugar, proteins and

631 microbial product (Lavallee and others 2020), whereas cPOM and mPOM compartments were  
632 mostly constituted by highly-depleted lignin (Camino-Serrano and others 2019).

633 Finally, the lower SOC-A<sub>1</sub> beneath *S. richardsonii*, in conjunction with the higher soil bulk  
634  $\delta^{15}\text{N}$  with greater TLN<sub>w</sub> in the soil horizon A<sub>2</sub> (Figure 5e), suggests that rhizosphere priming  
635 might also contribute to the soil organic carbon dynamics in our sites. The increase in OM  
636 mineralization and related positive priming have been observed in many places across the  
637 Arctic (Hartley and others 2012; Keuper and others 2020). However, we did not quantify  
638 priming *per se*, and therefore additional field studies are now required to ascertain the link  
639 between plant nutrient demand traits and priming-driven soil carbon loss. In addition, analyses  
640 of carbon isotopes can involve potential sources of error, and we are aware that this study has  
641 some limitations that should be considered when placing the results in the geomorphological  
642 context of the valley. The studied sites are located along alluvial fans that allow for running  
643 water and deposition of sediments and carbon coming from the bordering plateau (Fortier and  
644 others 2006). Additional <sup>14</sup>C measurements across additional fluxes and pools would fine-tune  
645 the temporal dynamics of active and abandoned channels, and consequently that of *S.*  
646 *richardsonii* colonization.

## 647 **Conclusion**

648 Structural changes related to greater erect shrub abundance in the Arctic are strong regulators  
649 of soil organic carbon stocks through optimization of aboveground functional traits associated  
650 with carbon and nutrient acquisition dynamics. This study first highlights that the use of plant  
651 economic strategies instead of vegetation composition or plant functional groups may provide  
652 new insights on carbon cycling variation with vegetation changes. It remains to be tested to  
653 what extent other plant compartments such as roots and leaf litter contribute to differences in  
654 soil organic carbon stocks between vegetation conditions. Moreover, we evaluated soil organic  
655 carbon stocks along alluvial fans specifically, and complement in that way the handful of

656 studies that so far have provided soil carbon contents by accounting for the specificity of  
657 geomorphological terrain units in the Arctic. Lastly, fine-scale plant-environment interactions  
658 suggest that estimating soil organic carbon stocks locally may be difficult to extrapolate over  
659 larger areas, illustrating the need for carbon cycle studies in many more sites across the Arctic.  
660 In that context, establishing common functional trait-based methodologies that are easily  
661 applicable globally has the potential to foster our understanding of the consequences of  
662 vegetation changes on Arctic ecosystem carbon storage.

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961 **Table legends**

962 **Table 1.** Mean values ( $\pm 1$  SE) of response variables measured in plots outside and beneath  
963 *Salix richardsonii* (SR) patches along active and abandoned channels.  $n = 6$  per vegetation  
964 condition per channel, except for soil total water content ( $n = 12$ ) and soil temperature ( $n = 3$ ).  
965 Test statistics are for fixed effects from linear mixed models. *F*-values in bold indicate  
966 significant effects, which were adjusted with the FDR procedure (see Supplementary Material  
967 2). Measurement units are as follows: biomass in gDW m<sup>-2</sup> soil, LAI<sub>w</sub> in m<sup>2</sup> leaf m<sup>-2</sup> soil, TLN<sub>w</sub>  
968 in g m<sup>-2</sup> soil, leaf  $\delta^{13}\text{C}/\delta^{15}\text{N}$  in ‰, thaw front depth in cm, soil bulk density in cm<sup>3</sup>, soil total  
969 water content in %, soil temperature (FDD/TDD) in °C, soil total N content in kgN m<sup>-3</sup> soil,  
970 and volumetric soil organic carbon stocks (SOC) in kgC m<sup>-3</sup> soil.

971 **Table 2.** Results of linear mixed models testing the effect of leaf area index (LAI<sub>w</sub>) and total  
972 leaf nutrient content (TLN<sub>w</sub>) of woody plants (*Salix* spp.) on soil organic carbon stock. LAI<sub>w</sub>  
973 and TLN<sub>w</sub> were used as a proxy of plant carbon supply and nutrient demand, respectively.  
974 Predictors were scaled in the regression analysis. See Figure 4b,c for corresponding  
975 illustrations. Note that the difference between marginal and conditional  $r^2$  corresponds to  
976 random effects (pairs of quadrats nested within channels within fans).

**Table 1.**

	Active channels		Abandoned channels		Channel		Vegetation		Channel x veg.	
	Outside <i>SR</i>	Beneath <i>SR</i>	Outside <i>SR</i>	Beneath <i>SR</i>	<i>df</i>	<i>F</i>	<i>df</i>	<i>F</i>	<i>df</i>	<i>F</i>
<i>Plant traits</i>										
Total biomass	351.0 ± 93.2	1392.2 ± 250.4	308.1 ± 70.5	1504.4 ± 281.5	1	0.03	1	<b>32.19</b>	1	0.15
Aboveground biomass	112.5 ± 46.2	1007.2 ± 199.5	252.5 ± 61.5	1416.3 ± 207.5	1	3.15	1	<b>51.75</b>	1	0.88
Root biomass	481.5 ± 181.5	780.1 ± 121.4	538.7 ± 133.4	670.5 ± 172.7	1	0.02	1	2.63	1	0.39
Shrub aboveground biomass	99.6 ± 45.6	1004.8 ± 200.0	41.1 ± 7.2	1240.2 ± 223.6	1	0.33	1	<b>48.57</b>	1	0.97
Graminoid aboveground biomass	12.9 ± 6.6	2.4 ± 1.3	3.2 ± 1.7	1.4 ± 1.2	1	1.66	1	<b>4.88</b>	1	2.41
Moss biomass	0.0 ± 0.0	0.0 ± 0.0	211.4 ± 57.4	174.7 ± 44.0	1	<b>28.66</b>	1	0.26	1	0.26
Shrub leaf biomass	35.6 ± 13.0	120.7 ± 23.1	20.7 ± 3.8	58.2 ± 25.0	1	4.14	1	<b>10.36</b>	1	1.56
Shrub stem biomass	64.0 ± 20.7	884.1 ± 185.9	17.2 ± 5.7	1182.0 ± 207.3	1	0.80	1	<b>50.64</b>	1	1.53
Leaf area index (LAI <sub>w</sub> )	0.3 ± 0.1	1.0 ± 0.1	0.4 ± 0.1	0.7 ± 0.1	1	1.31	1	<b>52.89</b>	1	<b>9.51</b>
Total leaf nutrient content (TLN <sub>w</sub> )	13.3 ± 5.8	51.3 ± 10.0	6.7 ± 1.7	26.3 ± 11.6	1	3.63	1	<b>12.17</b>	1	1.24
Leaf δ <sup>13</sup> C	-28.9 ± 0.3	-29.6 ± 0.3	-28.7 ± 0.1	-29.1 ± 0.3	1	1.90	1	2.98	1	0.18
Leaf δ <sup>15</sup> N	-6.7 ± 0.7	-5.5 ± 0.7	-6.1 ± 0.4	-6.0 ± 0.4	1	0.01	1	3.82	1	2.57
<i>Soil conditions</i>										
Thaw front depth	6.9 ± 0.1	7.6 ± 0.5	8.8 ± 0.8	8.7 ± 0.6	1	3.10	1	198.00	1	2.99
Horizon A <sub>1</sub> depth	1.1 ± 0.3	1.7 ± 0.4	2.2 ± 0.2	2.4 ± 0.3	1	5.70	1	3.70	1	1.03
Horizon A <sub>2</sub> depth	5.8 ± 0.2	5.9 ± 0.1	5.9 ± 0.2	5.6 ± 0.3	1	0.30	1	0.25	1	2.13
Soil bulk density	0.9 ± 0.1	0.8 ± 0.1	0.8 ± 0.1	0.8 ± 0.1	1	0.43	1	0.09	1	0.07
Total water content	3.2 ± 0.3	3.1 ± 0.2	4.5 ± 0.4	4.7 ± 0.5	1	<b>9.10</b>	1	0.12	1	0.03
pH	7.6 ± 0.1	7.5 ± 0.1	7.1 ± 0.1	7.2 ± 0.1	1	<b>9.57</b>	1	0.67	1	5.16
Freezing degree-days (FDD)	5114.6 ± 220.6	4968.9 ± 114.9	6080.8 ± 174.6	6637.1 ± 428.2	1	<b>25.17</b>	1	0.61	1	1.79
Thawing degree-days (TDD)	298.0 ± 53.4	285.3 ± 67.5	223.9 ± 48.4	112.6 ± 31.3	1	<b>5.30</b>	1	1.34	1	0.85
Soil C:N ratio	9.0 ± 0.7	8.6 ± 0.3	9.0 ± 0.6	10.4 ± 0.5	1	2.36	1	1.06	1	3.23
Soil total N content	1.4 ± 0.2	2.2 ± 0.2	2.0 ± 0.2	1.7 ± 0.1	1	0.23	1	1.33	1	7.16
<i>Soil organic carbon stocks</i>										
Volumetric SOC <sub>tot</sub>	15.62 ± 1.22	19.04 ± 1.12	18.82 ± 0.86	18.49 ± 1.03	1	1.17	1	3.09	1	4.55
Volumetric SOC-A <sub>1</sub>	34.58 ± 4.69	27.30 ± 2.70	30.59 ± 2.15	27.37 ± 2.10	1	0.25	1	7.43	1	1.11
Volumetric SOC-A <sub>2</sub>	12.40 ± 1.60	16.68 ± 1.87	16.24 ± 1.28	15.51 ± 0.96	1	0.65	1	1.99	1	3.98
Volumetric C-cPOM	0.73 ± 0.22	1.06 ± 0.05	1.15 ± 0.24	1.41 ± 0.31	1	2.29	1	2.43	1	0.04
Volumetric C-mPOM	0.75 ± 0.19	1.57 ± 0.31	1.38 ± 0.18	1.44 ± 0.31	1	0.85	1	3.53	1	2.71
Volumetric C-fPOM	0.61 ± 0.11	1.52 ± 0.27	1.06 ± 0.17	1.28 ± 0.26	1	0.26	1	7.26	1	2.70
Volumetric C-cPOM-A <sub>1</sub>	3.03 ± 0.72	3.60 ± 0.76	3.08 ± 0.39	3.63 ± 0.82	1	0.00	1	0.75	1	0.00
Volumetric C-cPOM-A <sub>2</sub>	0.40 ± 0.21	0.58 ± 0.07	0.54 ± 0.22	0.41 ± 0.07	1	0.01	1	0.03	1	1.11
Volumetric C-mPOM-A <sub>1</sub>	5.94 ± 1.58	5.55 ± 0.58	5.51 ± 0.85	5.11 ± 1.03	1	0.14	1	0.17	1	0.00
Volumetric C-mPOM-A <sub>2</sub>	0.05 ± 0.01	0.55 ± 0.16	0.12 ± 0.04	0.07 ± 0.01	1	6.12	1	7.49	1	10.64
Volumetric C-fPOM-A <sub>1</sub>	4.79 ± 1.32	4.59 ± 0.48	3.91 ± 0.81	4.42 ± 0.98	1	0.26	1	0.03	1	0.17
Volumetric C-fPOM-A <sub>2</sub>	0.08 ± 0.02	0.53 ± 0.17	0.22 ± 0.04	0.15 ± 0.02	1	1.69	1	5.45	1	9.93

**Table 2.**

	Soil organic carbon stock (kg C m <sup>-3</sup> soil)		
	Estimate	CI	<i>P</i>
Predictors			
Intercept	18.0	(16.5, 19.5)	< 0.001
LAI <sub>w</sub>	2.44	(1.36,3.52)	< 0.001
TLN <sub>w</sub>	-1.79	(-2.95, -0.64)	0.002
Random effects			
σ <sup>2</sup>	3.93		
τ <sub>00</sub> pair:(channel:fan)	0.00		
τ <sub>00</sub> channel :fan	0.71		
τ <sub>00</sub> fan	0.42		
Marginal <i>r</i> <sup>2</sup> / Conditional <i>r</i> <sup>2</sup>			0.47 / 0.51

## Figure legends

**Figure 1.** Conceptual model showing the direction of the expected interactions between variables in the studied system. The following paths were considered: (1) *Salix richardsonii* increases directly soil organic carbon stock, independently from plant traits. (2) Plants supply organic carbon to the soil, which increases soil organic carbon (SOC) stock. (3) Plants enhance N-rich SOC mineralization for their nutritional requirements, which not only releases mineral nutrients in the soil but also reduces SOC stock. (4) Plant C:N stoichiometry is maintained via a strong coordination between plant carbon supply and nutrient demand. The occurrence of the erect shrub *S. richardsonii* increases (5) carbon supply and (6) nutrient demand, compared to adjacent plots dominated by graminoid and prostrate shrub species. (7) Active channels provide water and nutrients to the surrounding vegetation, which stimulates plant growth and increases plant carbon supply to the soil; but they may also disturb plant standing biomass, which would decrease plant carbon supply. (8) Active channels provide water to the surrounding vegetation, which stimulates transpiration and nutrient mass-flow uptake, thus decreasing plant nutrient demand; (9) A shift in hydrological regime influence directly soil organic carbon stock, with higher organic carbon stored in soils with higher fertility, i.e. located along active channels.

**Figure 2.** Study location and experimental design. (a) The study was conducted in the Qarlikturvik valley of Bylot Island, Nunavut (blue circle and red rectangle in the left panel, respectively), along two alluvial fans (white rectangle in the middle panel). The white arrows in the right panel indicate the water flow going down from the plateau to the valley floor. (b) Along each fan, plant and soil materials were sampled in two hydrological regimes corresponding to active and abandoned channels (full and dotted white arrows, respectively), and where *Salix richardsonii* (*SR*) has settled. (c) In each channel zone, a paired sampling was carried out using plots where *S. richardsonii* was either present (beneath *SR*) or absent (outside

SR). Soil cores, that were on average 8 cm deep, were sampled within 0.7 m x 0.7 m quadrats. A<sub>1</sub> and A<sub>2</sub> refer to the soil horizons sampled.

**Figure 3.** Volumetric soil organic carbon (SOC, kg m<sup>-3</sup>) outside and beneath *Salix richardsonii* (SR) patches located along active and abandoned channels. Horizontal lines refer to medians. Panel (a) refers to total SOC (SOC<sub>tot</sub>), while panels (b) and (c) represent SOC content per horizon and OM compartment ( $n = 6$  per vegetation condition per channel). The particulate organic matter compartments were discriminated as follows: coarse organic matter (cPOM, 2 mm to 1 mm), medium organic matter (mPOM, 1 mm to 250  $\mu$ m) and fine organic matter (fPOM, 250  $\mu$ m to 63  $\mu$ m). The mineral-associated OM compartment (MAOM), estimated as the difference between SOC<sub>tot</sub> and SOC of the three measured POM compartments, is represented for information purposes only. See Table 1 for corresponding statistics. † $P < 0.1$ .

**Figure 4.** Influence of vegetation on total soil organic carbon stock (SOC<sub>tot</sub>, kg m<sup>-3</sup> soil). (a) Importance of woody plant (*Salix* spp.; c.f. subscript 'w') traits and moss biomass on SOC<sub>tot</sub> variation, issued from an automated model selection analysis where the following explicative variables were considered: leaf area index (LAI<sub>w</sub>), specific leaf area (SLA<sub>w</sub>), total leaf nutrient (TLN<sub>w</sub>), leaf biomass (Leaf<sub>w</sub>), stem biomass (Stem<sub>w</sub>), leaf nutrient content (LNuC<sub>w</sub>), moss biomass, and root biomass (Root<sub>w</sub>). (b,c) Conditional relationships of the mixed regression model testing the impact of (c) leaf area index (LAI<sub>w</sub>) and (b) total leaf nutrient (TLN<sub>w</sub>) of woody plants (*Salix* spp.) on soil organic carbon stock (i.e. volumetric SOC content; see Table 2 for model statistics). Each panel considered the effect of the variation of one explicative variable, while the second variable was considered fixed at its median value; the y-axes refer to residual values of conditional relationships. (d) Best fitted path analysis model showing the pathways that were retained from the conceptual model (Figure 1). Blue and red arrows refer to positive and negative relations, respectively. Numbers refer to the coefficients of

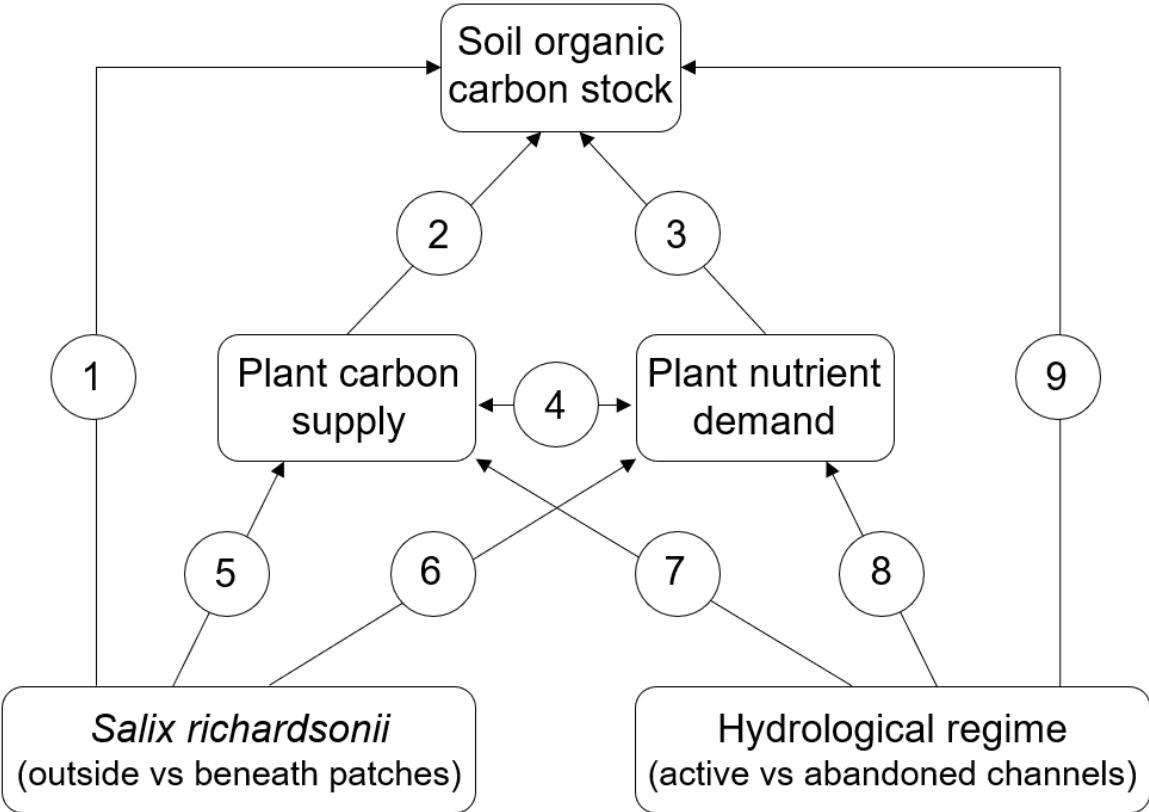
relationships between the predictor and response variables. Solid and dashed lines refer to significant and non-significant relations, respectively. LAI<sub>w</sub> and TLN<sub>w</sub> were considered as a proxy of plant net carbon supply and nutrient demand, respectively.

**Figure 5.** Influence of leaf area index (LAI<sub>w</sub>) and total leaf nutrient content (TLN<sub>w</sub>) of woody plants (*Salix* spp.) on soil organic carbon quality. LAI<sub>w</sub> and TLN<sub>w</sub> represent plant carbon supply and nutrient demand, respectively (see Figure 4). (a-c) Relationships between soil bulk  $\delta^{13}\text{C}$  and LAI<sub>w</sub>, determined according to (a) vegetation conditions and channel zones, (b) soil horizons, and (c) particulate OM compartments. (d-f) Relationships between soil bulk  $\delta^{15}\text{N}$  and TLN<sub>w</sub>, determined according to (d) vegetation conditions and channel zones, (e) soil horizons, and (c) particulate OM compartments of the soil horizon A<sub>2</sub>. Relationships per OM compartments of the top soil horizon A<sub>1</sub> were not drawn given that no relationship was previously observed for this given horizon (see panel e).

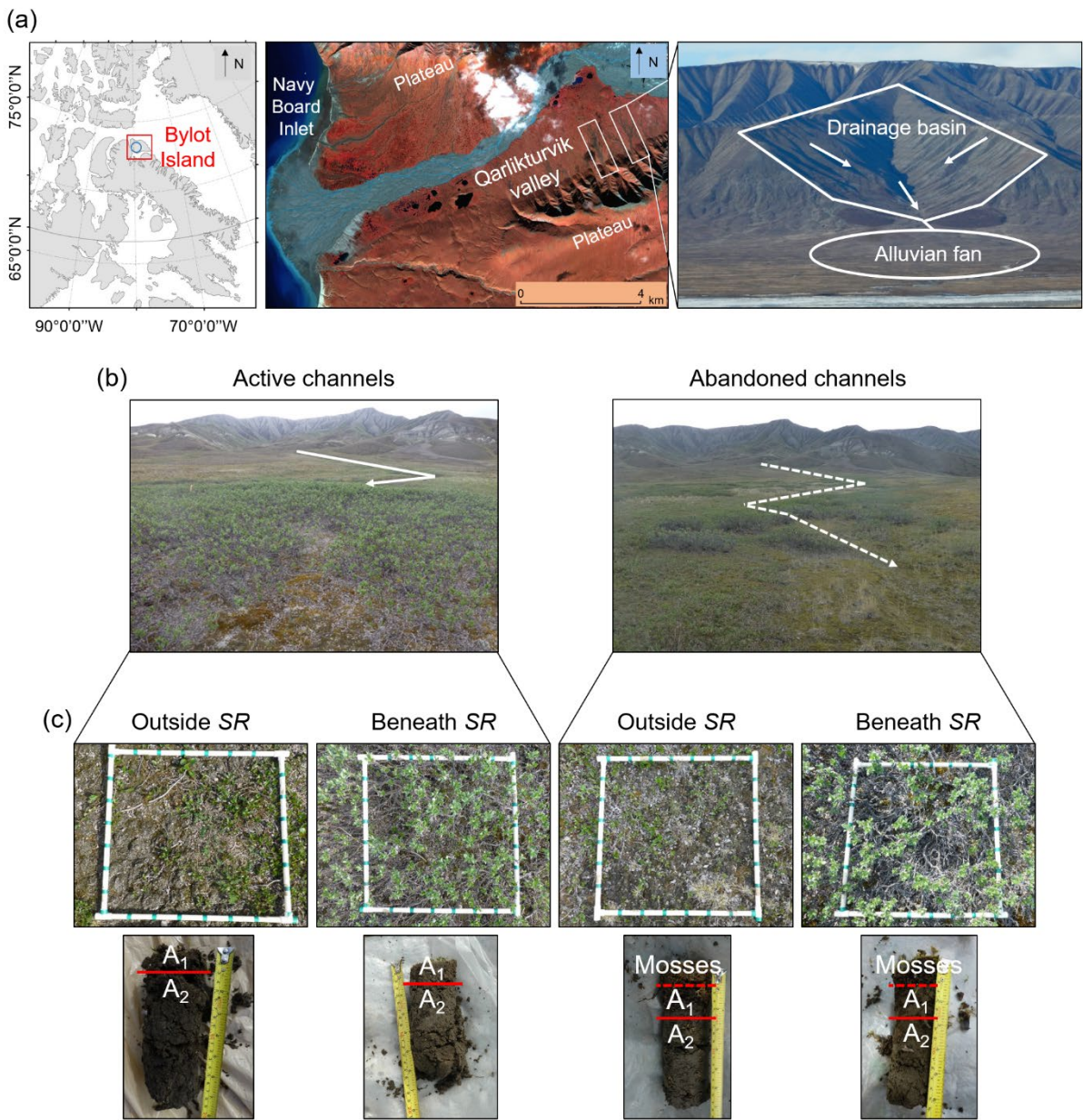
**Figure 6.** Total soil organic carbon stocks (SOC<sub>tot</sub>, kg m<sup>-3</sup>), as quantified per vegetation condition (outside and beneath *Salix richardsonii* (SR) patches) and particulate organic matter compartments (cPOM, mPOM and fPOM) of soil horizon A<sub>2</sub>, in relation to soil bulk  $^{14}\text{C}$  (composite samples from all repetitions). The particulate organic matter compartments were discriminated as follows: coarse organic matter (cPOM, 2 mm to 1 mm), medium organic matter (mPOM, 1 mm to 250  $\mu\text{m}$ ) and fine organic matter (fPOM, 250  $\mu\text{m}$  to 63  $\mu\text{m}$ ).



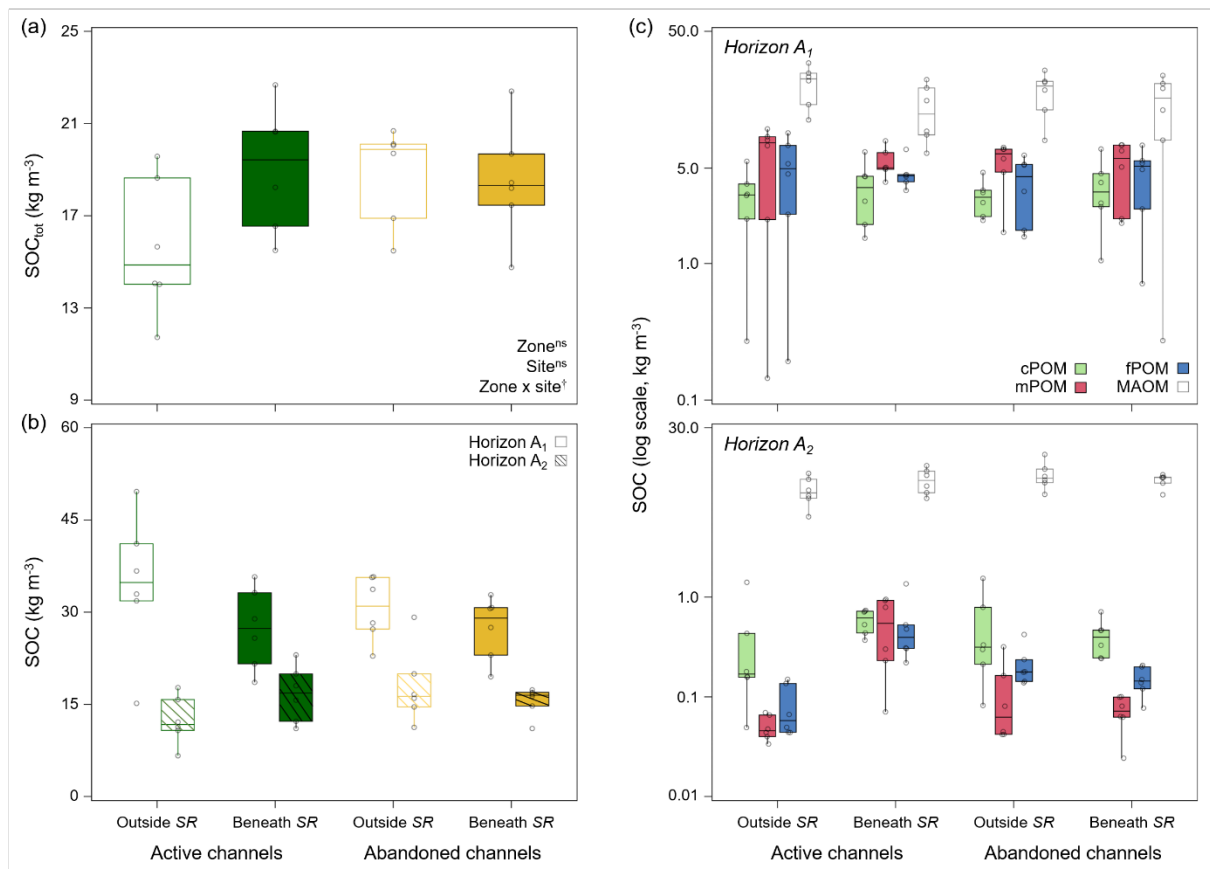
Figure 1



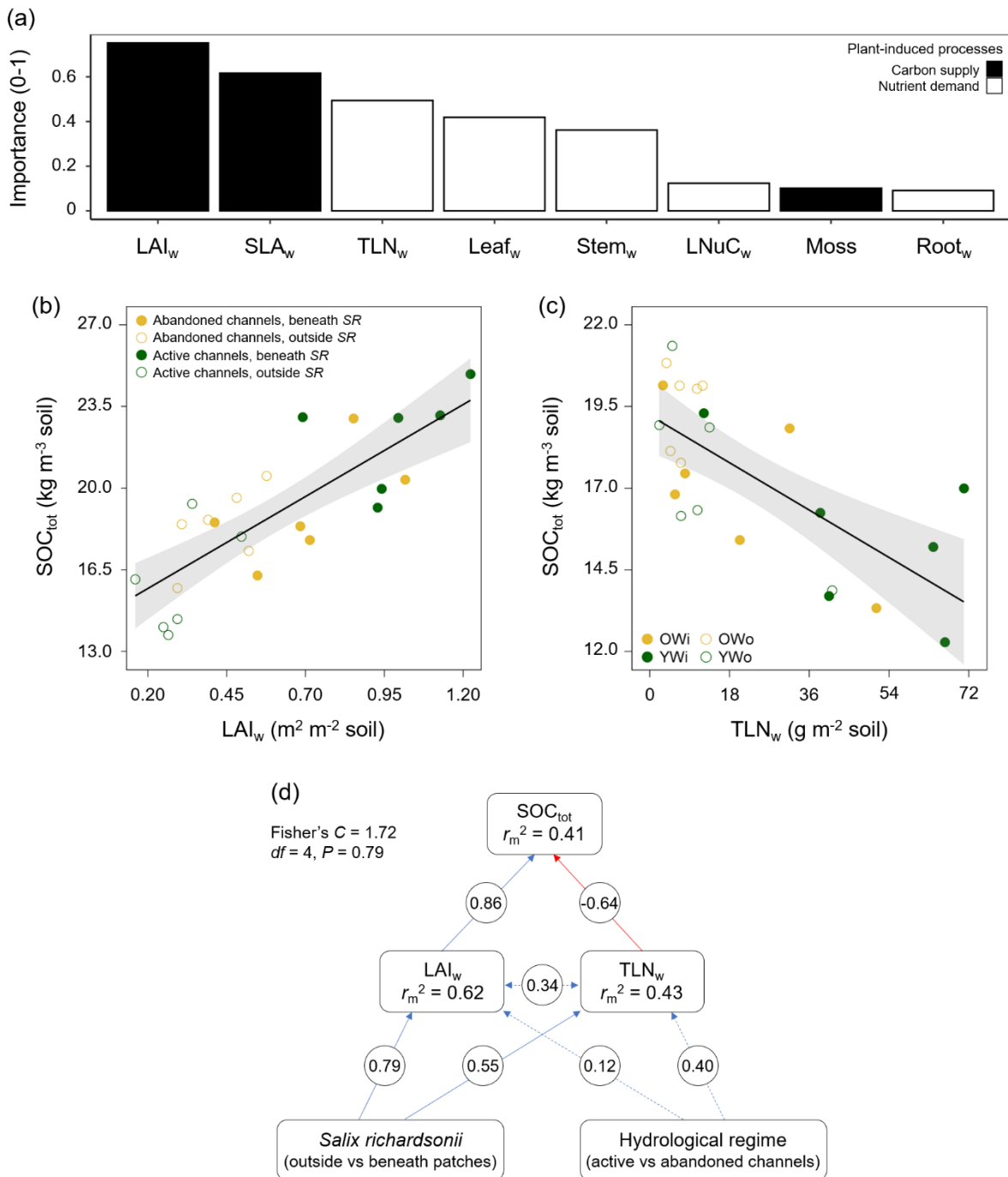
**Figure 2**



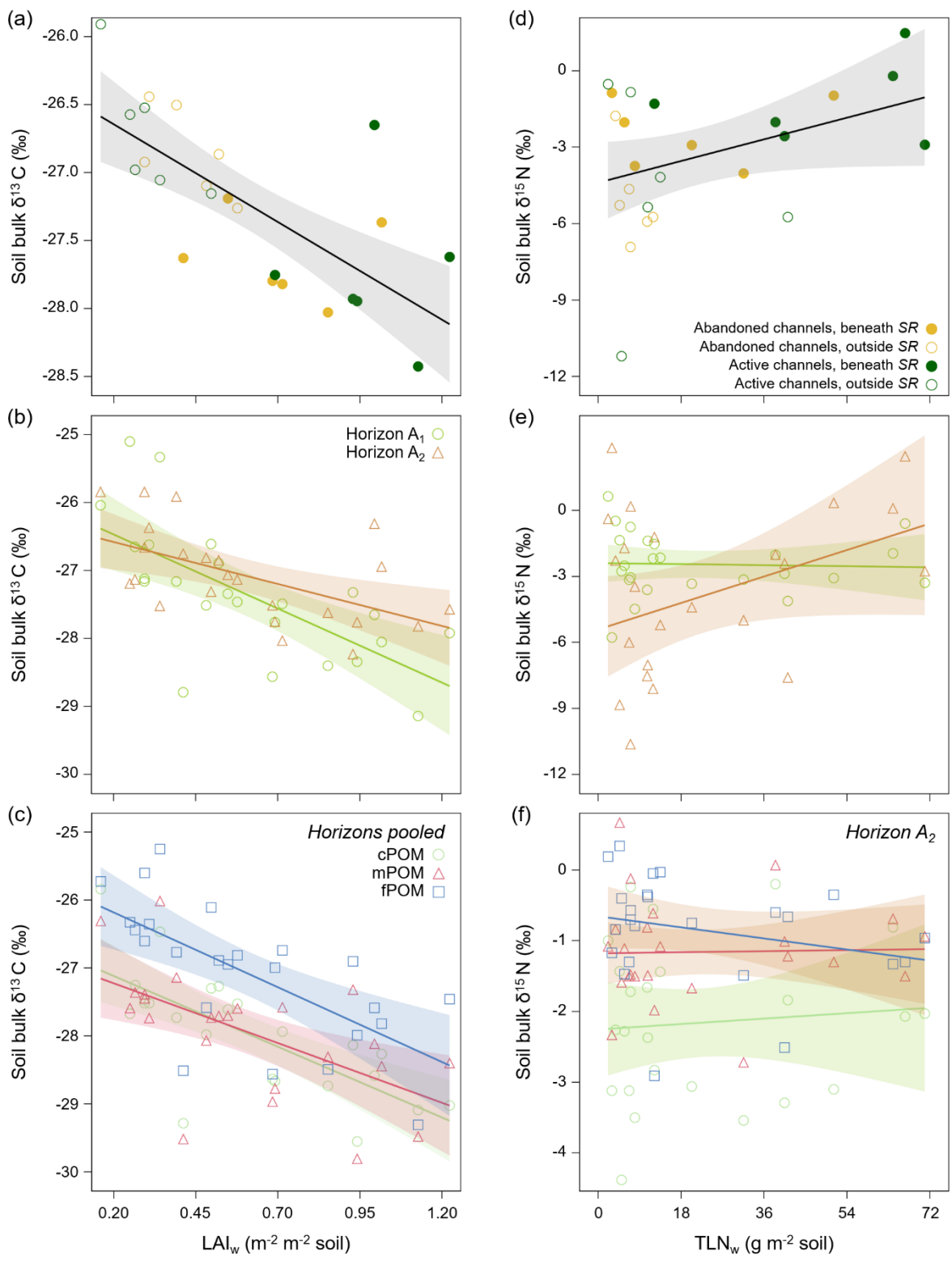
**Figure 3**



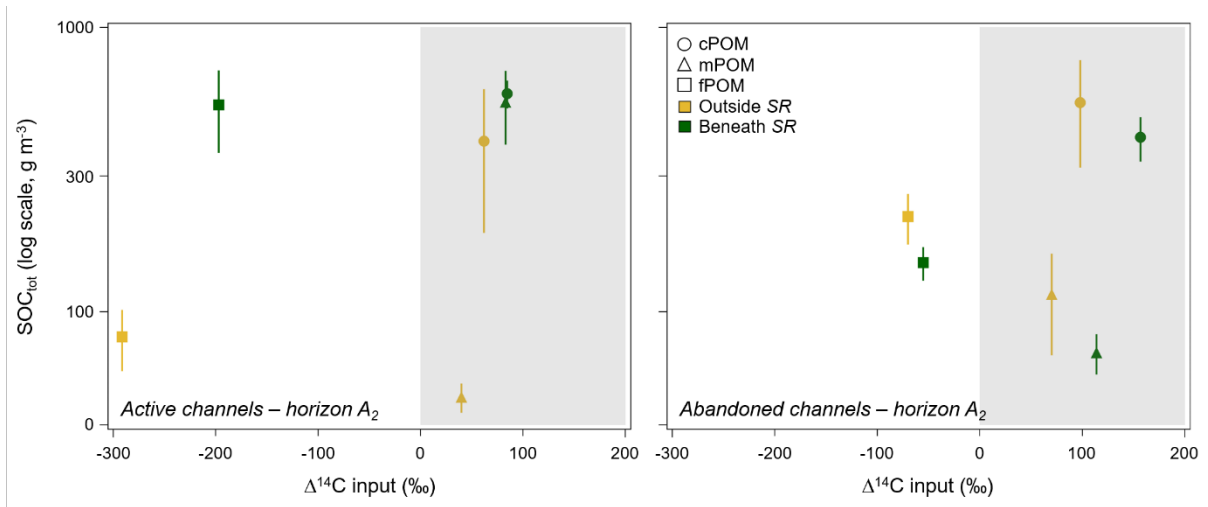
**Figure 4**



**Figure 5**



**Figure 6**



**Supplementary material 1.** Mean ( $\pm 1$  SE) values of response variables in plots located outside and beneath *Salix richardsonii* (SR) patches along active and abandoned channels.  $n = 6$  per vegetation condition per channel for each response variable. Test statistics are for fixed effects from linear mixed models.  $F$ -values in bold indicate significant effects, adjusted with the FDR procedure (see Supplementary Material 2). Measurement units are as follows: plant abundance in % cover, plant species richness in # species, leaf number in # leaves  $m^{-2}$ , leaf size in  $cm^2$  leaf $^{-1}$ ,  $SLA_w$  in  $m^2$  gDW $^{-1}$  leaf, leaf nutrient content in  $10^{-2}$  gNutrient gDW $^{-1}$ , leaf C:N in gC gN $^{-1}$ , root N content in  $10^{-2}$  gN gDW $^{-1}$ , surfacic (i.e. area based) soil organic carbon stocks (SOC) in kgC  $m^{-2}$  soil.

	Active channels		Abandoned channels		Channel		Vegetation		Channel x veg.	
	Outside SR	Beneath SR	Outside SR	Beneath SR	df	F	df	F	df	F
<i>Plant community</i>										
<i>Salix richardsonii</i> abundance	0.1 $\pm$ 0.1	54.2 $\pm$ 5.3	0.5 $\pm$ 0.4	66.3 $\pm$ 3.8	1	3.72	1	<b>341.79</b>	1	3.24
Graminoid abundance	0.0 $\pm$ 0.0	0.0 $\pm$ 0.0	0.1 $\pm$ 0.1	0.0 $\pm$ 0.0	1	0.96	1	1.04	1	0.96
Moss abundance	26.8 $\pm$ 15.6	9.6 $\pm$ 5.7	53.0 $\pm$ 14.8	32.5 $\pm$ 5.0	1	3.90	1	3.36	1	0.03
Species richness	5.5 $\pm$ 0.8	5.0 $\pm$ 1.0	6.5 $\pm$ 0.5	4.7 $\pm$ 0.7	1	0.14	1	3.68	1	1.20
<i>Plant traits</i>										
Leaf number	1313 $\pm$ 200	2821 $\pm$ 200	1546 $\pm$ 183	2475 $\pm$ 308	1	0.06	1	<b>28.58</b>	1	1.61
Leaf size	2.3 $\pm$ 0.0	3.5 $\pm$ 0.0	2.8 $\pm$ 0.1	2.9 $\pm$ 0.0	1	0.94	1	<b>90.63</b>	1	<b>79.11</b>
Specific leaf area ( $SLA_w$ )	75.7 $\pm$ 0.5	67.2 $\pm$ 0.2	71.1 $\pm$ 1.3	66.7 $\pm$ 0.0	1	12.26	1	<b>84.57</b>	1	8.05
Leaf C content	45.9 $\pm$ 0.2	44.5 $\pm$ 0.4	46.2 $\pm$ 0.3	44.3 $\pm$ 0.3	1	0.02	1	<b>76.93</b>	1	2.14
Leaf N content	2.8 $\pm$ 0.1	3.2 $\pm$ 0.1	2.6 $\pm$ 0.1	3.1 $\pm$ 0.1	1	1.53	1	<b>18.57</b>	1	0.65
Leaf P content	4.1 $\pm$ 0.3	5.5 $\pm$ 0.3	4.8 $\pm$ 0.5	6.1 $\pm$ 0.5	1	2.28	1	<b>11.44</b>	1	0.03
Leaf K content	16.3 $\pm$ 0.7	17.2 $\pm$ 0.7	16.0 $\pm$ 1.0	17.2 $\pm$ 1.1	1	0.03	1	1.24	1	0.03
Leaf Ca content	10.8 $\pm$ 0.8	12.7 $\pm$ 0.6	9.3 $\pm$ 0.6	12.7 $\pm$ 0.7	1	0.76	1	<b>38.66</b>	1	3.14
Leaf Mg content	3.7 $\pm$ 0.2	4.2 $\pm$ 0.1	3.8 $\pm$ 0.1	4.4 $\pm$ 0.4	1	0.25	1	<b>9.18</b>	1	0.08
Leaf nutrient content (LNuC <sub>w</sub> )	37.8 $\pm$ 1.4	42.7 $\pm$ 1.4	36.5 $\pm$ 1.4	43.4 $\pm$ 1.4	1	0.05	1	<b>17.58</b>	1	0.48
Leaf C:N ratio	16.4 $\pm$ 0.7	14.2 $\pm$ 0.5	17.6 $\pm$ 0.3	14.3 $\pm$ 0.5	1	1.63	1	<b>27.89</b>	1	1.17
Root N content	1.3 $\pm$ 0.2	1.4 $\pm$ 0.1	1.7 $\pm$ 0.1	1.3 $\pm$ 0.1	1	1.02	1	1.57	1	3.89
<i>Soil organic carbon stocks</i>										
Surfacic SOC <sub>tot</sub>	1.07 $\pm$ 0.08	1.48 $\pm$ 0.15	1.64 $\pm$ 0.06	1.55 $\pm$ 0.03	1	9.37	1	4.62	1	<b>11.36</b>
Surfacic SOC-A <sub>1</sub>	0.31 $\pm$ 0.02	0.44 $\pm$ 0.11	0.66 $\pm$ 0.06	0.63 $\pm$ 0.05	1	12.47	1	0.85	1	1.77
Surfacic SOC-A <sub>2</sub>	0.73 $\pm$ 0.10	0.99 $\pm$ 0.12	0.95 $\pm$ 0.07	0.87 $\pm$ 0.07	1	0.22	1	1.46	1	4.55
Surfacic C-cPOM	0.05 $\pm$ 0.02	0.08 $\pm$ 0.01	0.10 $\pm$ 0.02	0.12 $\pm$ 0.03	1	4.98	1	2.13	1	0.11
Surfacic C-mPOM	0.05 $\pm$ 0.01	0.13 $\pm$ 0.03	0.12 $\pm$ 0.02	0.13 $\pm$ 0.03	1	1.79	1	4.15	1	2.57
Surfacic C-fPOM	0.04 $\pm$ 0.01	0.12 $\pm$ 0.03	0.09 $\pm$ 0.02	0.11 $\pm$ 0.03	1	0.84	1	6.18	1	2.47
Surfacic C-cPOM-A <sub>1</sub>	0.03 $\pm$ 0.01	0.05 $\pm$ 0.00	0.07 $\pm$ 0.01	0.10 $\pm$ 0.03	1	6.45	1	3.05	1	0.04
Surfacic C-cPOM-A <sub>2</sub>	0.02 $\pm$ 0.01	0.03 $\pm$ 0.00	0.03 $\pm$ 0.01	0.02 $\pm$ 0.00	1	0.07	1	0.04	1	1.02
Surfacic C-mPOM-A <sub>1</sub>	0.05 $\pm$ 0.01	0.09 $\pm$ 0.02	0.11 $\pm$ 0.02	0.13 $\pm$ 0.03	1	3.18	1	2.54	1	0.88
Surfacic C-mPOM-A <sub>2</sub>	0.00 $\pm$ 0.00	0.03 $\pm$ 0.01	0.01 $\pm$ 0.00	0.00 $\pm$ 0.00	1	6.27	1	7.37	1	<b>10.61</b>
Surfacic C-fPOM-A <sub>1</sub>	0.04 $\pm$ 0.01	0.09 $\pm$ 0.03	0.08 $\pm$ 0.02	0.10 $\pm$ 0.03	1	1.79	1	3.44	1	0.47
Surfacic C-fPOM-A <sub>2</sub>	0.00 $\pm$ 0.00	0.03 $\pm$ 0.01	0.01 $\pm$ 0.00	0.01 $\pm$ 0.00	1	1.92	1	5.25	1	<b>9.39</b>

**Supplementary material 2.** False Discovery Rate (FDR) procedure applied on variables used in linear mixed models testing the effect of hydrological regime and *Salix richardsonii* (SR) occurrence on plant communities and soil conditions (c.f. Table 1 and Supplementary material 1). The procedure was applied separately for each of the fixed factors, i.e. channel (active vs abandoned), vegetation condition (outside vs beneath *S. richardsonii*), and the interaction between channel and vegetation condition. Values in bold indicate significant effects after the FDR procedure was applied. In the FDR procedure, the significant threshold is set at the highest  $P$ -value at which the inequality  $P \leq q$  holds true, with this and all smaller  $P$ -values corresponding to significant comparisons (Benjamini and Hochberg 1995; but see Pike 2011 for the method applied in ecology and evolution). Here,  $q = (P_{\text{rank}}/\text{number of tests}) * 0.05$ .

	Channel				Vegetation				Channel x vegetation		
	$P$	rank	$q$		$P$	rank	$q$		$P$	rank	$q$
<i>Plant community</i>				<i>Plant community</i>				<i>Plant community</i>			
SR abundance	0.083	1	0.013	<b>SR abundance</b>	<b>&lt;.0001</b>	<b>1</b>	<b>0.013</b>	SR abundance	0.102	1	0.013
Moss abundance	0.077	2	0.025	Species richness	0.084	2	0.025	Species richness	0.298	2	0.025
Graminoid abundance	0.350	3	0.038	Moss abundance	0.097	3	0.038	Graminoid abundance	0.350	3	0.038
Species richness	0.718	4	0.050	Graminoid abundance	0.332	4	0.050	Moss abundance	0.874	4	0.050
<i>Plant traits</i>				<i>Plant traits</i>				<i>Plant traits</i>			
<b>Moss biomass</b>	<b>0.0003</b>	<b>1</b>	<b>0.002</b>	<b>Total abg biomass</b>	<b>&lt;.0001</b>	<b>1</b>	<b>0.002</b>	<b>Leaf size</b>	<b>&lt;.0001</b>	<b>1</b>	<b>0.002</b>
SLA <sub>w</sub>	0.006	2	0.004	<b>Shrub abg biomass</b>	<b>&lt;.0001</b>	<b>2</b>	<b>0.004</b>	<b>LAI<sub>w</sub></b>	<b>0.001</b>	<b>2</b>	<b>0.004</b>
Shrub leaf biomass	0.069	3	0.006	<b>Shrub shoot biomass</b>	<b>&lt;.0001</b>	<b>3</b>	<b>0.006</b>	SLA <sub>w</sub>	0.018	3	0.006
TLN <sub>w</sub>	0.086	4	0.008	<b>LAI<sub>w</sub></b>	<b>&lt;.0001</b>	<b>4</b>	<b>0.008</b>	Root N content	0.077	4	0.008
Total abg biomass	0.106	5	0.010	<b>Leaf size</b>	<b>&lt;.0001</b>	<b>5</b>	<b>0.010</b>	Leaf Ca content	0.107	5	0.010
Leaf P content	0.162	6	0.013	<b>SLA<sub>w</sub></b>	<b>&lt;.0001</b>	<b>6</b>	<b>0.013</b>	Leaf δ <sup>15</sup> N	0.140	6	0.013
Leaf δ <sup>13</sup> C	0.198	7	0.015	<b>Leaf C content</b>	<b>&lt;.0001</b>	<b>7</b>	<b>0.015</b>	Graminoid abg biomass	0.152	7	0.015
Graminoid abg biomass	0.227	8	0.017	<b>Leaf Ca content</b>	<b>&lt;.0001</b>	<b>8</b>	<b>0.017</b>	Leaf C content	0.174	8	0.017
Leaf C:N ratio	0.231	9	0.019	<b>Total plant biomass</b>	<b>0.0002</b>	<b>9</b>	<b>0.019</b>	Leaf number	0.233	9	0.019
Leaf N content	0.244	10	0.021	<b>Leaf number</b>	<b>0.0003</b>	<b>10</b>	<b>0.021</b>	Shrub leaf biomass	0.240	10	0.021
LAI <sub>w</sub>	0.278	11	0.023	<b>Leaf C:N ratio</b>	<b>0.0004</b>	<b>11</b>	<b>0.023</b>	Shrub shoot biomass	0.245	11	0.023
Root N content	0.336	12	0.025	<b>Leaf N content</b>	<b>0.002</b>	<b>12</b>	<b>0.025</b>	TLN <sub>w</sub>	0.291	12	0.025
Leaf size	0.355	13	0.027	<b>LNuC<sub>w</sub></b>	<b>0.002</b>	<b>13</b>	<b>0.027</b>	Leaf C:N ratio	0.306	13	0.027
Shrub shoot biomass	0.393	14	0.029	<b>TLN<sub>w</sub></b>	<b>0.006</b>	<b>14</b>	<b>0.029</b>	Shrub abg biomass	0.349	14	0.029
Leaf Ca content	0.403	15	0.031	<b>Leaf P content</b>	<b>0.007</b>	<b>15</b>	<b>0.031</b>	Total abg biomass	0.369	15	0.031
Shrub abg biomass	0.581	16	0.033	<b>Shrub leaf biomass</b>	<b>0.009</b>	<b>16</b>	<b>0.033</b>	Leaf N content	0.438	16	0.033
Leaf Mg content	0.625	17	0.035	<b>Leaf Mg content</b>	<b>0.013</b>	<b>17</b>	<b>0.035</b>	LNuC <sub>w</sub>	0.506	17	0.035
Leaf number	0.811	18	0.038	Graminoid abg biomass	0.052	18	0.038	Root biomass	0.544	18	0.038
LNuC <sub>w</sub>	0.834	19	0.040	Leaf δ <sup>15</sup> N	0.079	19	0.040	Moss biomass	0.621	19	0.040
Leaf K content	0.857	20	0.042	Leaf δ <sup>13</sup> C	0.115	20	0.042	Leaf δ <sup>13</sup> C	0.679	20	0.042
Total plant biomass	0.864	21	0.044	Root biomass	0.136	21	0.044	Total plant biomass	0.702	21	0.044
Root biomass	0.883	22	0.046	Root N content	0.239	22	0.046	Leaf Mg content	0.780	22	0.046
Leaf C content	0.901	23	0.048	Leaf K content	0.292	23	0.048	Leaf K content	0.857	23	0.048



Leaf $\delta^{15}\text{N}$	0.939	24	0.050		Moss biomass	0.621	24	0.050		Leaf P content	0.864	24	0.050
<i>Soil conditions</i>					<i>Soil conditions</i>					<i>Soil conditions</i>			
<b>FDD</b>	<b>0.001</b>	<b>1</b>	<b>0.005</b>		Horizon A <sub>1</sub> depth	0.083	1	0.005		Soil total N content	0.023	1	0.005
<b>pH</b>	<b>0.011</b>	<b>2</b>	<b>0.010</b>		Thaw front depth	0.190	2	0.010		pH	0.047	2	0.010
<b>Total water content</b>	<b>0.013</b>	<b>3</b>	<b>0.015</b>		TDD	0.262	3	0.015		Soil C:N ratio	0.102	3	0.015
<b>TDD</b>	<b>0.020</b>	<b>4</b>	<b>0.020</b>		Soil total N content	0.276	4	0.020		Thaw front depth	0.115	4	0.020
Horizon A <sub>1</sub> depth	0.038	5	0.025		Soil C:N ratio	0.327	5	0.025		Horizon A <sub>2</sub> depth	0.175	5	0.025
Thaw front depth	0.109	6	0.030		pH	0.433	6	0.030		FDD	0.218	6	0.030
Soil C:N ratio	0.155	7	0.035		FDD	0.457	7	0.035		Horizon A <sub>1</sub> depth	0.335	7	0.035
Soil bulk density	0.527	8	0.040		Horizon A <sub>2</sub> depth	0.626	8	0.040		TDD	0.369	8	0.040
Horizon A <sub>2</sub> depth	0.595	9	0.045		Total water content	0.733	9	0.045		Soil bulk density	0.801	9	0.045
Soil total N content	0.642	10	0.050		Soil water content	0.774	10	0.050		Total water content	0.857	10	0.050
<i>Volumetric SOC stocks</i>					<i>Volumetric SOC stocks</i>					<i>Volumetric SOC stocks</i>			
C-mPOM-A <sub>2</sub>	0.033	1	0.004		C-mPOM-A <sub>2</sub>	0.021	1	0.004		C-mPOM-A <sub>2</sub>	0.009	1	0.004
C-cPOM	0.161	2	0.008		SOC-A <sub>1</sub>	0.021	2	0.008		C-fPOM-A <sub>2</sub>	0.010	2	0.008
C-fPOM-A <sub>2</sub>	0.223	3	0.013		C-fPOM	0.023	3	0.013		SOC <sub>tot</sub>	0.059	3	0.013
SOC <sub>tot</sub>	0.305	4	0.017		C-fPOM-A <sub>2</sub>	0.042	4	0.017		SOC-A <sub>2</sub>	0.074	4	0.017
C-mPOM	0.378	5	0.021		C-mPOM	0.090	5	0.021		C-mPOM	0.131	5	0.021
SOC-A <sub>2</sub>	0.437	6	0.025		SOC <sub>tot</sub>	0.110	6	0.025		C-fPOM	0.131	6	0.025
C-fPOM	0.623	7	0.029		C-cPOM	0.150	7	0.029		SOC-A <sub>1</sub>	0.317	7	0.029
C-fPOM-A <sub>1</sub>	0.624	8	0.033		SOC-A <sub>2</sub>	0.189	8	0.033		C-cPOM-A <sub>2</sub>	0.341	8	0.033
SOC-A <sub>1</sub>	0.628	9	0.038		C-cPOM-A <sub>1</sub>	0.406	9	0.038		C-fPOM-A <sub>1</sub>	0.686	9	0.038
C-mPOM-A <sub>1</sub>	0.725	10	0.042		C-mPOM-A <sub>1</sub>	0.688	10	0.042		C-cPOM	0.843	10	0.042
C-cPOM-A <sub>2</sub>	0.931	11	0.046		C-fPOM-A <sub>1</sub>	0.860	11	0.046		C-cPOM-A <sub>1</sub>	0.990	11	0.046
C-cPOM-A <sub>1</sub>	0.953	12	0.050		C-cPOM-A <sub>2</sub>	0.871	12	0.050		C-mPOM-A <sub>1</sub>	0.997	12	0.050
<i>Surfacic SOC stocks</i>					<i>Surfacic SOC stocks</i>					<i>Surfacic SOC stocks</i>			
SOC-A <sub>1</sub>	0.005	1	0.004		C-mPOM-A <sub>2</sub>	0.022	1	0.004		<b>SOC<sub>tot</sub></b>	<b>0.007</b>	<b>1</b>	<b>0.004</b>
SOC <sub>tot</sub>	0.012	2	0.008		C-fPOM	0.032	2	0.008		<b>C-mPOM-A<sub>2</sub></b>	<b>0.009</b>	<b>2</b>	<b>0.008</b>
C-cPOM-A <sub>1</sub>	0.029	3	0.013		C-fPOM-A <sub>2</sub>	0.045	3	0.013		<b>C-fPOM-A<sub>2</sub></b>	<b>0.012</b>	<b>3</b>	<b>0.013</b>
C-mPOM-A <sub>2</sub>	0.031	4	0.017		SOC <sub>tot</sub>	0.057	4	0.017		SOC-A <sub>2</sub>	0.059	4	0.017
C-cPOM	0.050	5	0.021		C-mPOM	0.069	5	0.021		C-mPOM	0.140	5	0.021
C-mPOM-A <sub>1</sub>	0.105	6	0.025		C-fPOM-A <sub>1</sub>	0.093	6	0.025		C-fPOM	0.147	6	0.025
C-fPOM-A <sub>2</sub>	0.196	7	0.029		C-cPOM-A <sub>1</sub>	0.111	7	0.029		SOC-A <sub>1</sub>	0.213	7	0.029
C-mPOM	0.210	8	0.033		C-mPOM-A <sub>1</sub>	0.142	8	0.033		C-cPOM-A <sub>2</sub>	0.336	8	0.033
C-fPOM-A <sub>1</sub>	0.211	9	0.038		C-cPOM	0.175	9	0.038		C-mPOM-A <sub>1</sub>	0.371	9	0.038
C-fPOM	0.380	10	0.042		SOC-A <sub>2</sub>	0.255	10	0.042		C-fPOM-A <sub>1</sub>	0.508	10	0.042
SOC-A <sub>2</sub>	0.648	11	0.046		SOC-A <sub>1</sub>	0.378	11	0.046		C-cPOM	0.747	11	0.046
C-cPOM-A <sub>2</sub>	0.793	12	0.050		C-cPOM-A <sub>2</sub>	0.842	12	0.050		C-cPOM-A <sub>1</sub>	0.844	12	0.050

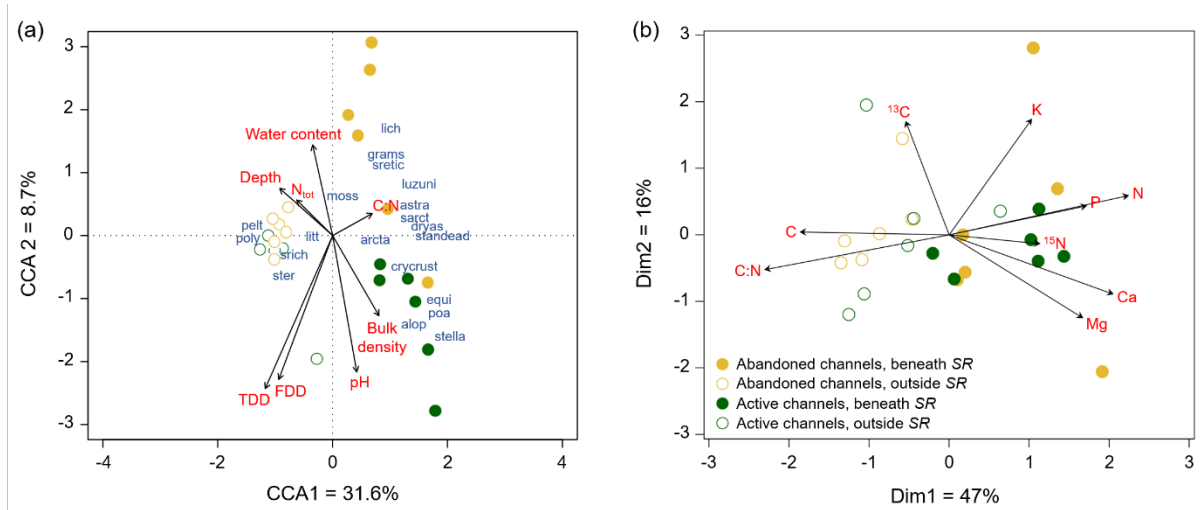
## Reference

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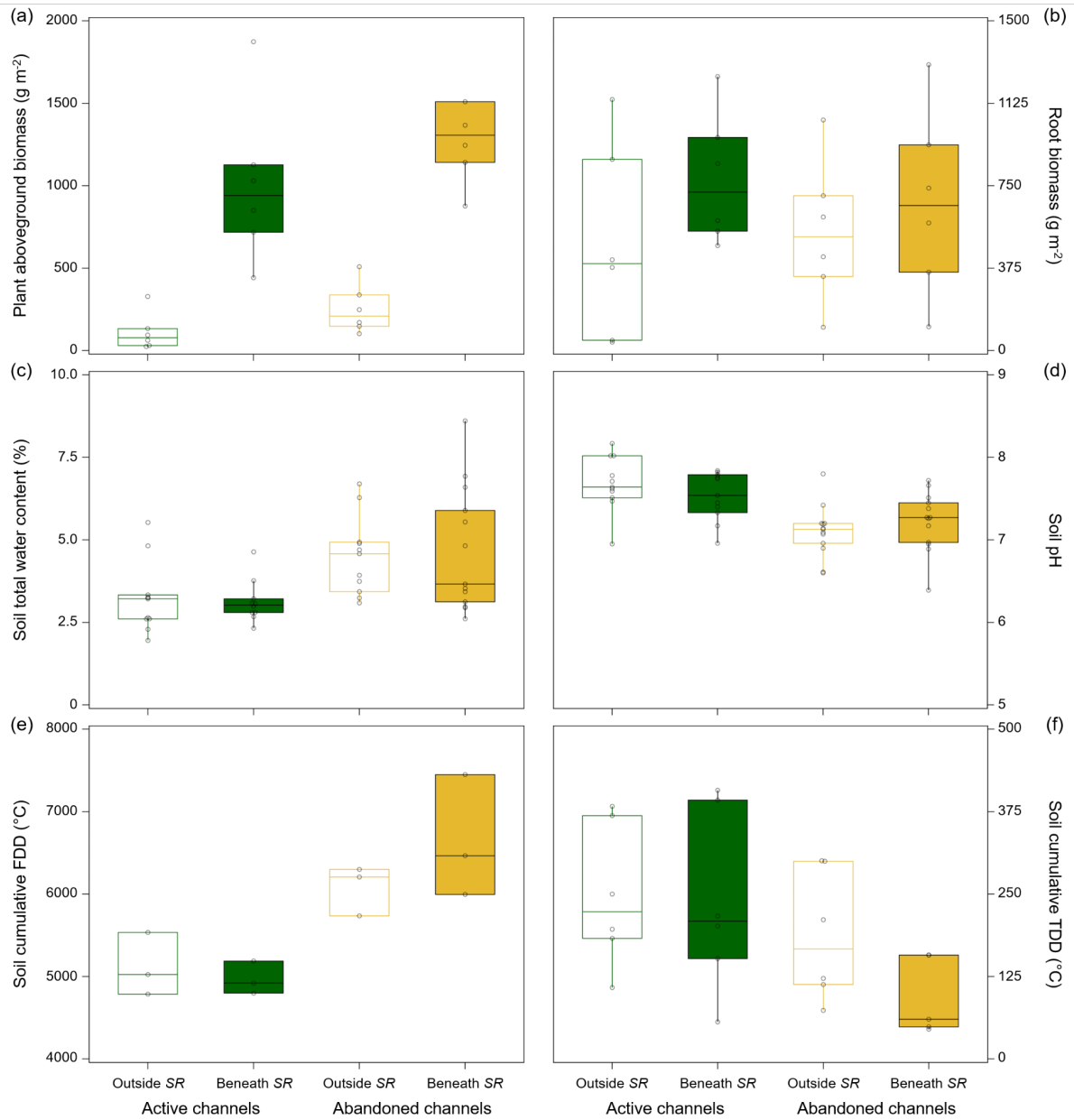
**Supplementary material 3.** Mean species richness and mean species cover of vascular and non-vascular taxa found in plots located outside and beneath *Salix richardsonii* (SR) patches along active and abandoned channels. Species richness and species abundance were estimated within a 0.7 m x 0.7 m quadrat arranged in each of the 24 sampled vegetation plots. < for cover < 0.1% and << for cover < 0.01%. Species names are reported according to the Database of Vascular Plants of Canada (VASCAN; <https://data.canadensys.net/vascan/search>).

	Active channels		Abandoned channels	
	Outside SR	Beneath SR	Outside SR	Beneath SR
Mean species richness	6	5	7	5
Plant species cover (%)				
Caryophyllaceae				
<i>Stellaria longipes</i> Goldie	<<	0	0	0
Equisetaceae				
<i>Equisetum arvense</i> L.	0.6	<<	<	<<
Fabaceae				
<i>Astragalus alpinus</i> L.	0.4	<<	1.8	<<
Juncaceae				
<i>Luzula nivalis</i> Spreng.	<<	0	0.4	0
Poaceae				
<i>Alopecurus magellanicus</i> Lam.	<	0	0	0
<i>Arctagrostis latifolia</i> (R. Br.) Griseb.	0.5	<	0.2	<
<i>Poa arctica</i> R. Br.	<<	0	0	0
Other spp.	<<	0	<	0
Polygonaceae				
<i>Bistorta vivipara</i> (L.) Delarbre	<<	<	<<	<<
Rosaceae				
<i>Dryas integrifolia</i> Vahl	0.5	<	0.9	0
Salicaceae				
<i>Salix richardsonii</i> Hooker	<	54.2	0.5	66.3
<i>Salix arctica</i> Pall.	3.3	<<	5.8	0.2
<i>Salix reticulata</i> L.	0.6	0	2.2	<
Lichens				
<i>Peltigera</i> spp.	0	0.4	0	0.4
<i>Stereocaulon</i> spp.	0	0.4	0	1.2
Other spp.	0	0	0.5	0
Mosses	26.8	9.6	53.0	32.5
Cryptogamic crust cover (%)	64.6	6.3	25.0	0
Litter cover (%)	3.1	41.7	5.8	45.8
Standing dead cover (%)	0.5	0	0.6	0

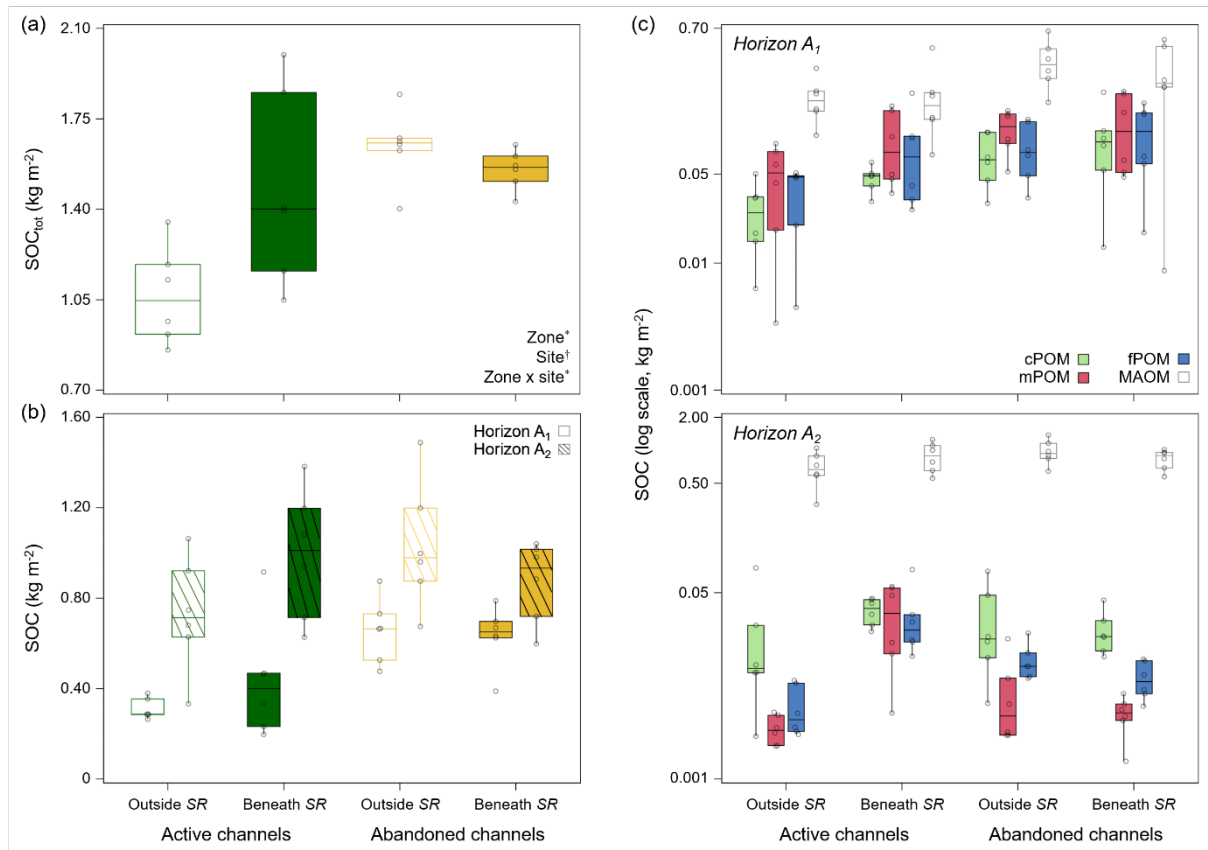
**Supplementary material 4.** (a) Canonical correspondence analysis testing the association between plant community and soil conditions (i.e. thaw front depth, bulk density, water content, pH, freezing degree-days, thawing degree-days, C:N ratio, and total N content). (b) Principal component analysis of plots located outside and beneath *Salix richardsonii* (SR) patches in active and abandoned channels, relative to leaf nutrients (i.e. C/N/P/K/Ca/Mg content, C:N ratio,  $^{13}\text{C}$  and  $^{15}\text{N}$  composition) of *Salix* spp.



**Supplementary material 5.** Plant biomass (a,b) and soil conditions (c-f) in plots located outside and beneath *Salix richardsonii* (SR) patches along active and abandoned channels.  $n = 6$  per vegetation condition per channel for each response variable, except for soil total water content ( $n = 12$ ) and soil freezing/thawing degree-days (FDD/TDD;  $n = 3$ ).



**Supplementary material 6.** Surfacic (i.e. area based) soil organic carbon (SOC, kg m<sup>-2</sup>) outside and beneath *Salix richardsonii* (SR) patches located along active and abandoned channels. Panel (a) refers to total SOC (SOC<sub>tot</sub>), while panels (b) and (c) represent SOC content per horizon and OM compartment ( $n = 6$  per vegetation condition per channel). The particulate organic matter compartments were discriminated as follows: coarse organic matter (cPOM, 2 mm to 1 mm), medium organic matter (mPOM, 1 mm to 250 μm) and fine organic matter (fPOM, 250 μm to 63 μm). The mineral-associated OM compartment (MAOM), estimated as the difference between SOC<sub>tot</sub> and SOC of the three measured POM compartments, is also represented. See Supplementary material 1 for corresponding statistics. \* $P < 0.05$ , † $P < 0.1$ .



**Supplementary material 7.**  $\delta^{13}\text{C}$ ,  $\delta^{15}\text{N}$  and C:N ratio of plant leaves, plant roots and OM compartments found in soil horizons  $A_1$  and  $A_2$  sampled in plots located outside and beneath *Salix richardsonii* (SR) patches along active and abandoned channels. The particulate organic matter compartments were discriminated as follows: coarse organic matter (cPOM, 2 mm to 1 mm), medium organic matter (mPOM, 1 mm to 250  $\mu\text{m}$ ) and fine organic matter (fPOM, 250  $\mu\text{m}$  to 63  $\mu\text{m}$ ).

