

Original Article

Validation protocol for assessing the upper cervical spine kinematics and helical axis: An *in vivo* preliminary analysis for axial rotation, modeling, and motion representation

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Abstract

Context: The function of the upper cervical spine (UCS) is essential in the kinematics of the whole cervical spine. Specific motion patterns are described at the UCS during head motions to compensate coupled motions occurring at the lower cervical segments. **Aims:** First, two methods for computing *in vitro* UCS discrete motions were compared to assess three-dimensional (3D) kinematics. Secondly, the same protocol was applied to assess the feasibility of the procedure for *in vivo* settings. Also, this study attempts to expose the use of anatomical modeling for motion representation including helical axis. **Settings and Design:** UCS motions were assessed to verify the validity of *in vitro* 3D kinematics and to present an *in vivo* procedure for evaluating axial rotation. **Materials and Methods:** *In vitro* kinematics was sampled using a digitizing technique and computed tomography (CT) for assessing 3D motions during flexion extension and axial rotation. To evaluate the feasibility of this protocol *in vivo*, one asymptomatic volunteer performed an MRI kinematics evaluation of the UCS for axial rotation. Data processing allowed integrating data into UCS 3D models for motion representation, discrete joint behavior, and motion helical axis determination. **Results:** Good agreement was observed between the methods with angular displacement differences ranging from 1° to 1.5°. Helical axis data were comparable between both methods with axis orientation differences ranging from 3° to 6°. *In vivo* assessment of axial rotation showed coherent kinematics data compared to previous studies. Helical axis data were found to be similar between *in vitro* and *in vivo* evaluation. **Conclusions:** The present protocol confirms agreement of methods and exposes its feasibility to investigate *in vivo* UCS kinematics. Moreover, combining motion analysis, helical axis representation, and anatomical modeling, constitutes an innovative development to provide new insights for understanding motion behaviors of the UCS.

Key words: Helical axis, *in vivo* kinematics, modeling, motion representation, upper cervical spine, validation

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INTRODUCTION

Regarding the global kinematics of the cervical spine, the major role of the suboccipital spine (UCS) resides in the completion of specific movements to compensate coupled motions occurring at the lower cervical segments.

In the clinical evaluation of cervical spine function, motion impairments can be demonstrated as both quantitative and qualitative disturbances. Additionally, these kinematic features could be frequently accompanied by pain or discomfort.^[1,2]

Examination methods for assessing segmental motion of the cervical spine usually use medical imaging system exposing subject to ionizing radiation such as computed tomography (CT) or radiography.^[3-5] Recently, innovative procedures using magnetic resonance imaging (MRI) have succeeded in analyzing three-dimensional (3D) motions and morphology.^[6,7]

For 3D motion analysis, rotation axis orientation is also described to provide qualitative data for kinematics disorders assessment. With regard to the cervical spine, such data are mainly suggested for global head movement^[2,8,9] or segmental displacements of the lower cervical spine.^[10] Yet, only few studies have reported such data in two-dimensional (2D) for the UCS.^[11,12]

Despite representative data reported for the *in vivo* kinematics of the UCS, there is a paucity of studies reporting such data including helical axis computation and motion representation.

The purpose of this study is to validate a method for assessing UCS 3D motions and to evaluate the feasibility of such a procedure for *in vivo* conditions. Also, this study attempts to expose the use of anatomical modeling for motion representation including helical axis.

MATERIALS AND METHODS

Kinematics validation method

In the first part of this protocol, one unembalmed human specimen was processed based on a previously described experimental procedure related to UCS kinematics.^[13] After soft tissue dissection and prior to motion assessment, technical markers (TM = aluminum balls, 4 mm diameter) were pasted on the UCS bones (occiput (C0), atlas (C1), and axis (C2)). Discrete motions were considered using different UCS poses of flexion extension (FE) and axial rotation (AR). Validation was carried out using two different acquisition methods, a digitizing procedure using a 3D digitizer (FARO, B06/Rev 18; Technologies Inc; USA), and CT (Siemens SOMATOM, helical mode, reconstruction: slice thickness = 0.5 mm, interslice spacing = 1 mm, image data format = DICOM 3.0).

For each UCS attitude, TMs spatial locations were computed successively from 3D digitizing (TM_{dig}) and CT imaging (TM_{CT}), using a customized experimental jig providing similar control of joint displacements in each method.^[14] Five discrete poses were collected through the entire range of motion in each plane of interest.

Also, to obtain 3D models of all UCS bones (C0, C1, and C2), CT data were processed following a data segmentation procedure using dedicated software (Amira 3.0®, Germany).

In vivo assessment

To evaluate the feasibility of the above protocol, one asymptomatic volunteer (37 years) was selected to perform

an MRI kinematics assessment of the UCS for axial rotation (Philips Achieva 3T MRI, 3 Tesla, Philips Health Care, Best, The Netherlands, thickness = 1 mm, field of view = 160 mm, reconstruction pixel size 0.7×0.7 mm 2 averages). Motion evaluation was carried out according a method previously developed by Ishii *et al.*^[6] for five UCS discrete positions in AR from neutral position to maximal physiological active rotation with an intermediate attitude on both sides. The subject was in supine position with a head support. For assuring pure axial rotation, the head was held perpendicular to the MRI table under supervision of one operator using the Frankfurt plane as a reference. For each UCS attitude, MRI acquisition lasted 5 min approximately.

In addition, the volunteer had performed a CT scan evaluation during an earlier clinical routine assessment less than 2 years. The latter examination was not related with cervical complaints.

Imaging data (CT and MRI) were processed to create individual anatomical models using the above mentioned software. On each MRI model, anatomical landmarks were defined by virtual palpation as represented in Figure 1 to compute discrete kinematics data. Reproducibility of this method is described elsewhere reporting an average palpation error less than 1 mm.^[13] CT data was only used to provide accurate 3D UCS model for motion representation.

Data registration, kinematics and motion representation

To combine motion visualization of the collected bone segments and their associated discrete kinematics, a registration method was used as described in details previously.^[13,15] Registration was performed using a computer graphics environment, LHP FusionBox software (www.openmaf.org) for providing data fusion.

For each bone, motion was computed according its spatial position and orientation within the local anatomical coordinate system as defined earlier.^[13] Thus C0-C1 and C0-C2 motions were defined in

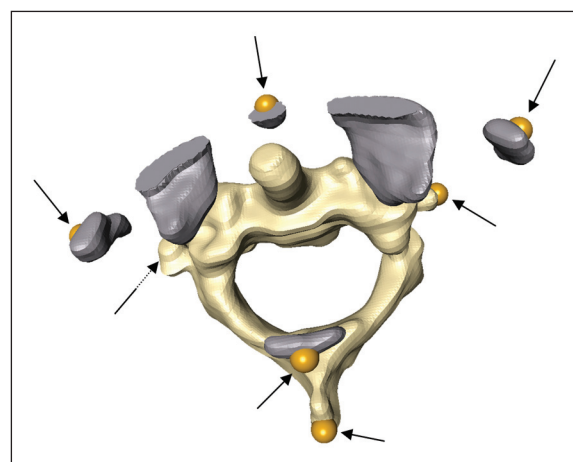


Figure 1: Three-dimensional (3D) atlantoaxial magnetic resonance imaging (MRI) model. Location of technical markers (arrows) following virtual palpation of anatomical landmarks. Model of C1 includes only necessary bone segments

the C1 and C2 reference systems, respectively. From the above data, finite helical axis (FHA) parameters (i.e., orientation and position) were determined and integrated into the 3D model to represent axis behavior over the range of motion for each UCS level.

RESULTS

Kinematics validation

For AR, global C0-C2 range of motion (ROM) was estimated to 52° and 53° according to CT and digitizing measurements, respectively. For FE, maximal amplitude reached 25° and 26°.

To evaluate agreement between the motion data, both methods were compared using estimation of absolute difference between measurements. Regarding discrete ROM, absolute mean differences were less than 1° for AR and ranged from 0.8° to 1.4° for FE. Discrete amplitudes are described in Figure 2 for each segment attitude. Concordance between angular displacements was demonstrated by coefficients of determination above 0.84 for rotation magnitudes superior to 1° [Table 1].

Concerning helical axis, orientation, and position were computed for both motions with an exception for C0-C1 AR due to the poor mobility at this level (5°). Absolute differences for FHA orientation ranged from 3.5° to 6.4° for C0-C1 FE and C1-C2 AR, respectively. Also for C1-C2 FE, only mean HA was considered due to limited global range of motion (8°). For the latter, orientation error was found to be less than 5°. As an example, Figure 3 illustrates FHA orientation and location for AR at C1-C2 level for both acquisition methods. Orientation of averaged FHA displayed a difference of 3° with a standard deviation (SD) of 1.4°

for both methods. For HA location, a maximal error of less than 1 mm was found regardless to anatomical reference axis.

In vivo assessment

Motion data (angular displacements and translations) are displayed in Table 2 related to AR at C1-C2. Principal angular displacement was observed around the Y-axis with a global magnitude of 71°. Coupled motions were demonstrated around secondary axes (X- and Z-axis) with maximal absolute rotation of 5° and 11°, respectively. For translations, small cephalocaudal and lateral displacements were observed. Figure 4 illustrated individual kinematics patterns occurring for angular displacements and translations throughout the entire motion range.

Averaged helical axis data (location and orientation) are presented in Table 3 and FHA graphical representation is displayed for three attitudes of C1-C2 AR [Figure 5]. Note the relative constant orientation of FHA aligned with the dens of C2, with an ipsilateral location to the direction of AR. Maximal FHA displacement was observed along the Z-axis with a magnitude of 11 mm for both rotation sides.

Table 1: Coefficients of determination (r^2) between computed tomography (CT) and digitizing kinematics data

	C0-C1			C1-C2		
	θX	θY	θZ	θX	θY	θZ
FE	0,972	0,919	0,990	0,213§	0,837	0,854
AR	0,982	0,011§	0,584§	0,969	0,999	0,302§

§ indicates discrete range of motion inferior to 1°

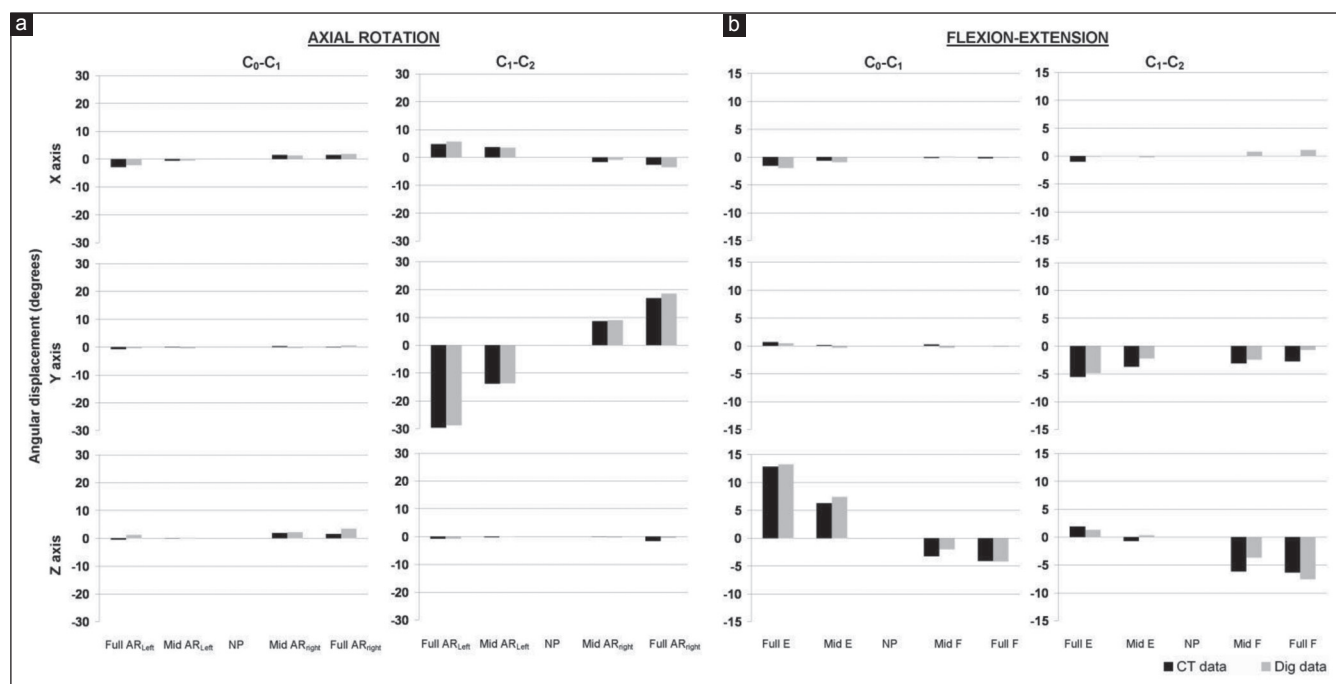


Figure 2: Kinematics data for axial rotation (AR) and (a) flexion extension (FE). (b) Helical angles computation. Representation of angular displacements of C0-C1 and C1-C2 for each discrete position for computed tomography (CT) and digitizing (Dig) measurements. NP=Neutral position

Table 2: *In vivo* kinematics data for C1-C2 axial rotation (AR)

	Helical angle (°)			Position (mm)		
	x	y	z	x	y	z
Right AR	-2,5	-37,7	-6,1	0,2	-3,1	-1,1
Left AR	4,9	33,7	10,6	-0,4	-1,1	1,1

Helical angles (°) and translation (mm) expressed in the anatomical reference system of C2

Table 3: Mean helical axis orientation (cosines) and position (mm)

	Orientation			Position		
	x	y	z	x	y	z
Mean	0,014	0,999	-0,043	-2,6	0,1	1,6
SD	0,004	0,000	0,009	0,9	0,2	3,7
max	0,020	0,999	-0,028	-0,9	0,3	5,6
min	0,007	0,998	-0,570	-4,1	-0,2	-5,0

Standard deviation (SD), minimal (min), and maximal (max) values

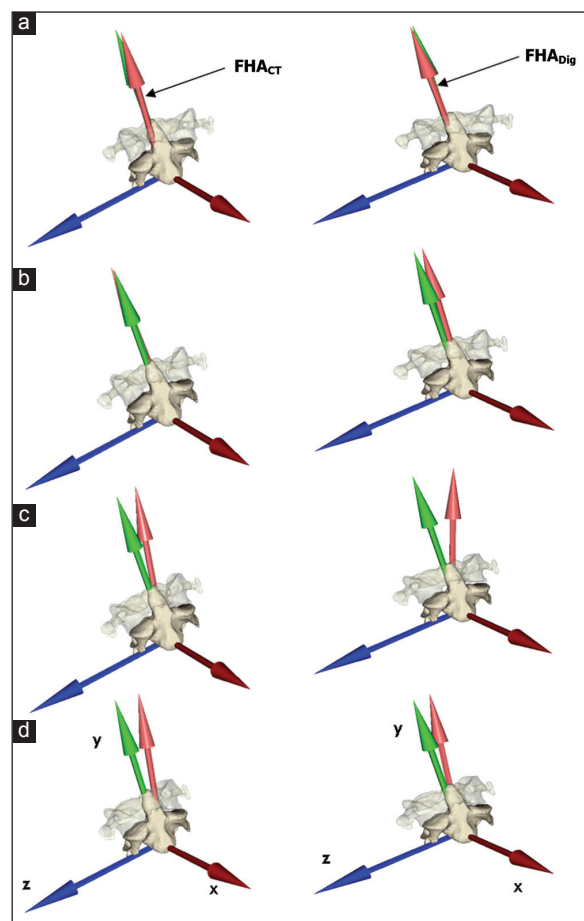


Figure 3: Representation of C1-C2 finite helical axis (FHA) location and orientation for digitizing (right column) and CT (left column) data between discrete motions (a, b, c, and d) from maximal right to left axial rotation, anterior views

DISCUSSION

The first part of the present study aimed at validating a procedure for assessing the kinematics of the UCS using two different methods for computing bone spatial location. This method is based on a previous study proposed by Van Sint Jan *et al.*,^[15] investigating elbow kinematics. The latter authors observed small motion divergences between comparable methods with maximal differences of about 1° for helical rotation. These results agreed with the present outcomes of rotation errors ranging from 1° to 1.5° depending on the level and the motion direction. Moreover, coefficients of determination demonstrated a good concordance between methods for helical angles for both motions of interest. Comparatively, earlier methods found angular errors ranging from 1° to 2° at the cervical spine either for global motion^[16] or for segmental motion.^[17,18]

Additionally, for helical axis orientation and position, the differences ranged from 3° to 6°, confirming good agreement between the methods for measuring UCS kinematics.

For the selected specimen, global ROM was found to be similar to previously reported *in vitro* data for AR but lesser for FE.^[13,19-23] However, for FE several authors reported similar values for *in vivo* conditions.^[18,24] For *in vitro* experimental methods, ROM discrepancies are generally recognized compared to that found for *in vivo* conditions. Moreover, lower

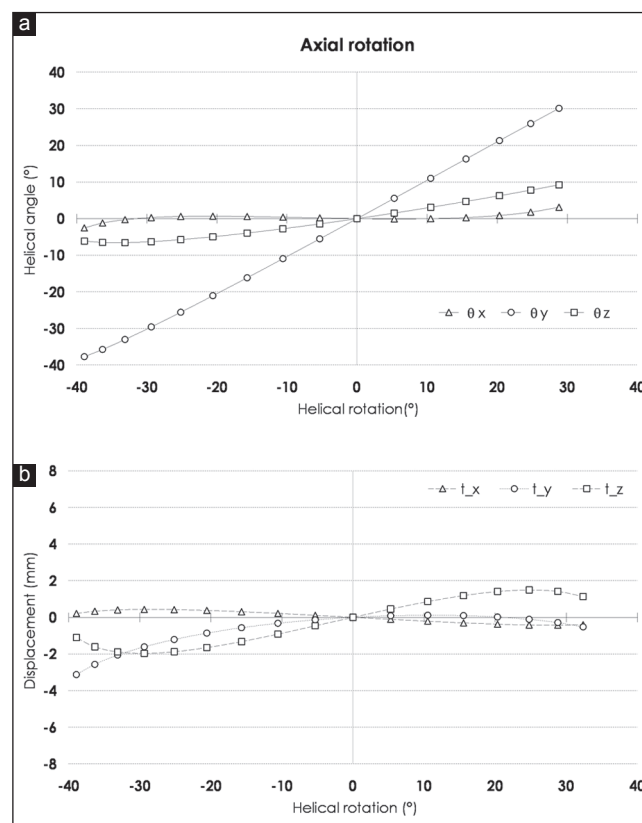


Figure 4: *In vivo* kinematics data, helical angles (θ in degrees) and (a) translations (t in mm) (b) for C1-C2 axial rotation (one single volunteer). Sample polynomial function (goodness of fit, $r^2 > 0.87$)

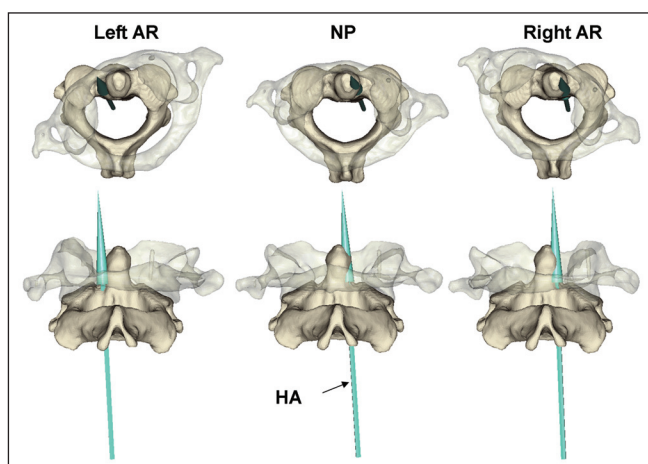


Figure 5: Atlantoaxial 3D-model and helical axis (HA) orientation and location during axial rotation *in vivo*. Superior view and posterior view for three discrete positions. NP=Neutral position

values could also be observed for some pathological conditions such as degenerative or rheumatoid processes at the UCS.^[7]

As mentioned in the literature, coupled motions frequently occur during AR or lateral bending of the UCS. These associated motion components are mainly reported as extension and heterolateral flexion during AR and as axial rotation (at C1-C2) for lateral bending. Here, coupled motions are mostly found at C1-C2 during flexion extension with a maximal magnitude around 5°. Such specific patterns have been mentioned earlier for FE^[20,25] and some authors linked these phenomena to the degenerative processes noticed at the UCS.^[7]

The second part of this work aimed at developing a noninvasive method for investigating the *in vivo* kinematics of the UCS using MRI acquisition. Based on a previously reported procedure, assessment was performed for five poses, from maximal left to right AR respecting the subject's comfort.^[6,26] Following this method, UCS axial rotation ROM represents 90% of the head rotation approximately.^[26] Kinematics was collected from MRI data performing virtual palpation of anatomical landmarks using specific computer graphic software. Such a method has already been recommended to provide spatial bone location as well as to build anatomical reference system. Using a registration method, kinematics and imaging data were fused to generate an anatomical model including kinematics behavior, HA motion representation as described earlier.^[13,15]

For our single volunteer, the global range of motion in AR was found to be similar to the data reported from studies relating *in vivo* assessment of the UCS with a value around 71°.^[5-7,26] Moreover, right rotation was slightly larger than left rotation, 37° versus 34°. This difference could be related either to the manual control of the subject's head position or to the compliance to rotational stress between upper and lower cervical spine.^[6]

Considering coupled motions, our results agreed with the recent data for *in vivo* conditions.^[6,26] In the present study, coupled lateral flexion was demonstrated in the opposite direction to AR

with an absolute magnitude ranging from 3° to 5°. For coupled motions in the sagittal plane, an irregular pattern was observed with a coupled extension during right rotation and a coupled flexion in left rotation, of 11° and 6°, respectively. As mentioned above, such motion pattern has been reported previously. Nevertheless, we must keep in mind that the lack of control of the subject's head position could be a source for asymmetric attitude in axial rotation^[5,6] as well as for the coupled motions. In this way, alternative methods were developed to standardize head attitude during such assessment.^[27,28]

During UCS motion, coupled translations are also reported as well for AR as for FE. These coupled motions are dependent on the measuring methods, but also on the reference system. In consideration for the latter, anatomical reference systems defined in this study are consistent with previous recommendations.^[20]

Similar to the validation data, analysis of the *in vivo* HA shows comparable orientation and location to previously reported data.^[13] As pointed out earlier, HA displays a vertical attitude trough to the dens of the axis during AR with moderate location variations depending on motion direction and subjects.^[5,13,29]

Thus, these singular outcomes show meaningful data regarding quantitative and qualitative *in vivo* kinematics compared to aforementioned *in vitro* data. Nevertheless, limitations of this study include the need of investigation for assessing reliability in a large sample before starting functional evaluation of UCS as clinical routine.

In conclusion, the protocol presented here confirms agreement of motion measurements and exposes its feasibility to investigate *in vivo* UCS kinematics. Moreover, combining motion analysis, helical axis representation and anatomical modeling, such innovative development provides new insights for understanding normal and abnormal motion behaviors of the UCS. Further investigations are now being started to integrate this method to evaluate other UCS motion types as well as the lower cervical spine kinematics.

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